

Development and evaluation of diverse promising rapeseed (*Brassica napus* L.) mutants using physical and chemical mutagens[☆]

Souhail Channaoui^{1,2}, Mostapha Labhilili³, Mohamed Mouhib⁴, Hamid Mazouz², Mohamed El Fectali¹ and Abdelghani Nabloussi^{1,*}

¹ Research Unit of Plant Breeding and Plant Genetic Resources Conservation, National Institute of Agricultural Research, Regional Agricultural Research Center of Meknes, Meknes, Morocco

² Laboratory of Plant Biotechnology and Molecular Biology, Department of Biology, Faculty of Science, University Moulay Ismail, Meknes, Morocco

³ Research Unit of Plant Biotechnology, National Institute of Agricultural Research, Regional Agricultural Research Center of Rabat, Rabat, Morocco

⁴ Research Unit of Nuclear Techniques, Environment and Quality, Regional Agricultural Research Center of Tangier, Tangier, Morocco

Received 5 February 2019 – Accepted 12 July 2019

Abstract – Genetic variability is a prerequisite for any plant breeding program, and mutagenesis is a proven way of creating new variation within a crop germplasm. Novel genetic variability in rapeseed was induced by gamma rays, Ethyl Methane Sulphonate (EMS) and combined mutagen treatment, using various doses and concentrations. The objective was to evaluate and compare the obtained M₂ mutants for important quantitative traits in two contrasted environments. Data on phenological, morphological and agronomic parameters were recorded. A large variability was observed and mutagenic treatments had a significant effect on all traits studied. Compared to control plants, mutant genotypes derived from seeds treated with low EMS concentrations during moderate time were earlier and characterized by a higher number of pods per plant. For high concentration of EMS during long time and for combinations of physical and chemical mutagens, a significant decrease in plant height and stature was noticed, as compared to control. Besides, plants derived from gamma rays-treated seeds exhibited the highest 1000-seed weight. The novel induced variability may be integrated in rapeseed breeding program as a new germplasm with improved agronomic traits. Particularly, EMS1-7-stable mutant may be exploited to develop efficiently and quickly a new rapeseed cultivar with some desirable traits. The present study highlights once more the possibility to bring novel genetic diversity for rapeseed desirable traits improvement through mutation breeding.

Keywords: rapeseed / quantitative traits / mutation breeding / EMS treatment / Gamma rays

Résumé – Développement et évaluation de divers mutants prometteurs de colza (*Brassica napus* L.) obtenus par mutagenèse physique et chimique. La variabilité génétique est essentielle pour tout programme d'amélioration de plantes et la mutagenèse s'avère une technique incontestable d'induction de nouvelle variabilité génétique dans les germoplasmes des cultures. Chez le colza, une nouvelle variabilité génétique a été induite après traitement de semences par les rayons gamma, l'EMS et leur combinaison, en utilisant différentes doses et concentrations. L'objectif de cette étude est d'évaluer et comparer les mutants M₂ obtenus pour des caractères quantitatifs, dans deux environnements contrastés. Les paramètres étudiés sont d'ordre phénologique, morphologique et agronomique. Une grande variabilité a été observée et les traitements mutagènes ont eu un effet significatif sur tous les paramètres. Les mutants qui proviennent d'une faible concentration d'EMS, durant une période modérée, sont plus précoces et ont un nombre de siliques plus élevé que la variété témoin (non traitée). Par contre, des mutants dérivés de forte concentration d'EMS, pendant une longue période, et de la combinaison d'EMS avec les rayons gamma, ont montré une réduction significative de la hauteur et de la vigueur des plantes. Par ailleurs, les plantes dérivées de

[☆] Contribution to the Topical Issue "Rapeseed / Colza"

*Correspondence: abdelghani.nabloussi@gmail.com

semences traitées aux rayons gamma ont eu le poids de 1000 graines le plus élevé. La nouvelle variabilité induite peut être intégrée et utilisée dans le programme de sélection du colza en tant que nouveau germplasm avec des caractéristiques agronomiques améliorées. En particulier, le mutant stable EMS1-7 peut être exploité pour développer efficacement et rapidement un nouveau cultivar de colza ayant des caractéristiques souhaitables. La présente étude indique une fois de plus la possibilité d'apporter une nouvelle diversité génétique pour l'amélioration des caractères d'intérêt du colza à travers la mutagenèse.

Mots clés : colza / caractères quantitatifs / amélioration par mutation / traitement EMS / rayons gamma

1 Introduction

Rapeseed (*Brassica napus* L.), an important oilseed crop, source of vegetable oil and protein-rich meal, is characterized by a substantially increased word production over the last 35 years, which currently reached six times the production recorded in 1980 (Wanasundara *et al.*, 2016). In 2016, the overall production was around 69 million tons (FAOSTAT, 2018). Rapeseed oil is mainly used in human nutrition and biofuel production, whereas rapeseed de-oiled meal is used in animal feed. In general, rapeseed oil contains ~7% saturated fatty acids (including palmitic acid and stearic acid), and high amounts of monounsaturated fatty acids with a significant fraction of oleic acid (~61%). It also contains an important amount of polyunsaturated fatty acids with significant fraction of linoleic acid (~11%) and α -linolenic acid (~21%) (Sharafi *et al.*, 2015).

In Morocco, as well as in other countries of Mediterranean area, rapeseed shows a good adaptation and has a great potential, as a promising oilseed crop that could play a role in improving the vegetable oils production in such countries. Thus, to enhance this production, there is a need to develop and release performant and adapted cultivars.

Available rapeseed germplasm naturally possesses limited genetic variability (Hasan *et al.*, 2006; Bus *et al.*, 2011). Consequently, conventional breeding methods are more and more restrictedly used to release the expected varieties (Tshilenge-Lukanda *et al.*, 2013). The use of mutagenesis to induce novel genetic variability is an effective approach for those crops with narrow genetic base such as rapeseed (Parry *et al.*, 2009). The main advantage of mutation breeding is the possibility of improving one or few characters without changing the rest of the genotype. In recent years, induced mutations have been extensively used for breeding annual oilseed crops (Spasibonek, 2006; Ferrie *et al.*, 2008; Velasco *et al.*, 2008; Emrani *et al.*, 2015; Hussain *et al.*, 2017). Mutagenesis has been also employed to improve a large number of desirable traits like as earliness, dwarfness, biotic and abiotic stress resistance or tolerance, seed yield and oil quality (Schnurbush *et al.*, 2000; Parry *et al.*, 2009; Ali and Shah, 2013; Lee *et al.*, 2018).

Many mutagen agents, either chemical or physical, are available to create and obtain valuable mutations in crop plants. Each particular mutagen agent acts according to a different and specific mode that determines the nature of alteration in plant genetic background (Meinke *et al.*, 1998). However, the biological effect of ionizing radiation like gamma rays depends primarily on the amount of energy that will be absorbed by the biological system for which the chromosomes are the most important target (Van Harten, 1998). Also, Ethyl Methane Sulphonate (EMS) is a chemical

mutagen of the alkylating group and has been commonly used in plant breeding because it can cause high frequency of gene mutations and low frequency of chromosome aberration (Van Harten, 1998). Both gamma rays and EMS have been successfully used in rapeseed to evolve new varieties with improved economic traits (Rahman, 1990; Shah *et al.*, 1999).

This study was carried out to compare the relative effectiveness of gamma rays and Ethyl Methane Sulphonate (EMS), applied alone and in combination, for inducing novel genetic variability in rapeseed, to evaluate the developed mutants, for some important traits, and to isolate and select mutants combining some desirable traits. Nevertheless, the study focused only on phenological and agronomic traits non related with seed quality.

2 Materials and methods

2.1 Plant material and treatments applied

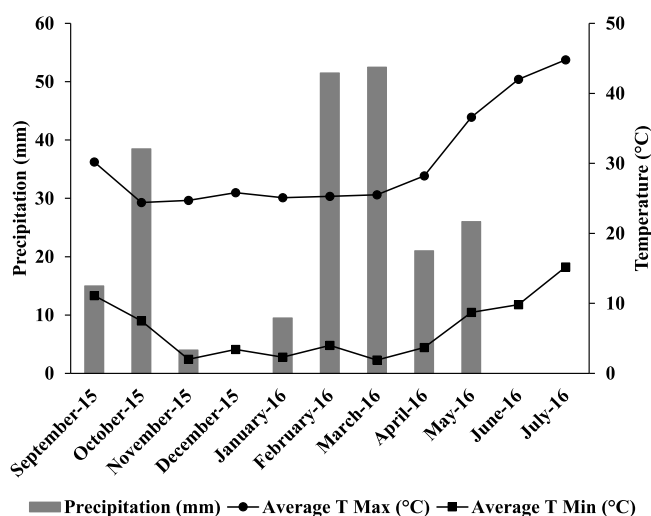
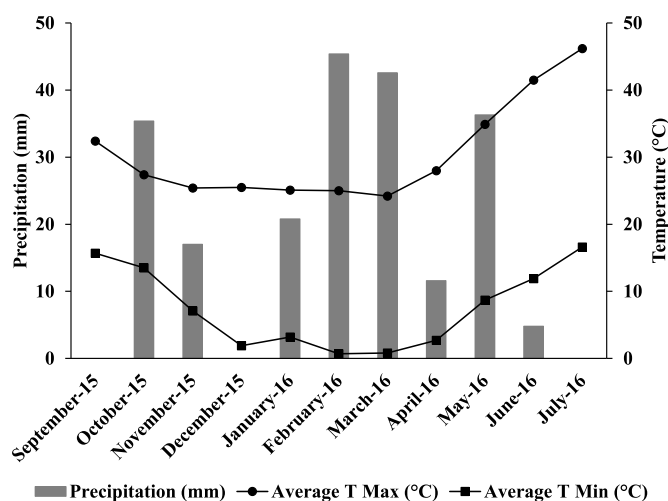
Plant material used in this study was the rapeseed (*Brassica napus* L.) variety "INRA-CZH2", from the collection of National Institute for Agricultural Research (INRA) of Morocco (Nabloussi, 2015). The physical mutagen (gamma rays) and chemical mutagen (Ethyl Methane Sulphonate, EMS) were applied for mutations induction. Seeds of this variety were subjected to one dose of gamma rays (1300 Gy), four different concentrations of EMS (1, 1.2, 1.4 and 1.6%) during 6, 7 and 14 h, and one treatment of these mutagens in combination (0.8% EMS during 6 h + 1100 Gy). Mutagen doses or concentrations used in the present study were designed on the basis of LD50 (lethal dose) results from seed germination test. DL50 is a dose that causes 50% mortality to the seeds, *i.e.* a safe dose where 50% of the seeds can survive. Many researchers think that a dose near to LD50 should be the optimum which varies with crop species and mutagen used (Singh, 2000). Thus, eleven treatment levels were considered in this study: One physical treatment by gamma rays, one combined treatment (EMS + gamma ray) and nine EMS treatments levels as shown in Table 1.

2.2 Field experiment

Treated (M_1) and control (M_0) seeds were sown on 5 m long-rows spaced 60 cm apart, according to a completely randomized design, without replications, on November 2014 at the INRA Experimental Station of Douyet (DYT). This station, at 416 m above the sea and with an average rainfall of 425 mm and cracking clay soil, is located at 10 km from Fez city (34°04' N, 5°07' W). Climate is of Mediterranean type, with cold and rainy winters, and hot and dry summers. This experimental station is also characterized by a frequent

Table 1. Concentrations/doses of mutagenic agents used to induce novel genetic variability in rapeseed.

Mutagen agents	Treatment	Treatment abbreviation
Gamma rays	1300 Gy	1300 Gy
Combined treatment	1100 Gy + 0.8% Concentration (%)	CombT
	1	EMS1-6
	1.2	EMS1.2-6
	1.4	EMS1.4-6
	1.6	EMS1.6-6
EMS	1	EMS1-7
	1.2	EMS1.2-7
	1	EMS1-14
	1.2	EMS1.2-14
	1.4	EMS1.4-14

**Fig. 1.** Average, maximum and minimum monthly temperatures and rainfall recorded in the Experimental Station of Douyet during 2015/2016.**Fig. 2.** Average, maximum and minimum monthly temperatures and rainfall recorded in the Experimental Station of Sidi Allal Tazi during 2015/2016.

presence of the sirocco wind which could be to some extent harmful for crop growing. At flowering, 20 individual M_1 plants, from each treatment, and exhibiting interesting attributes, regarding earliness and seed yield related traits, were selected and selfed to produce M_2 seeds.

Also, M_2 population was planted on 11 November 2015 at DYT and on 20 January 2016 at the INRA Experimental Station of Sidi Alal Tazi (ATZ), as a late sowing date. ATZ is located at 30 km from Kenitra city (34°31' N, 6° W), at an elevation of 10.5 m and with an average annual rainfall of 550 mm. The soil is limestone clay with higher salinity rate than DYT. Figures 1 and 2 show monthly temperatures and rainfall registered in DYT and ATZ, respectively, during the cropping season 2015/2016. In both locations, planting was done in a randomized complete block design on 5 m long rows spaced by 60 cm. Two rows of the check (untreated original variety seeds) were planted after every 10 rows of treated

material to facilitate the comparison during evaluation and selection.

During cropping cycle of M_2 population grown in DYT, minimum temperature was 2 °C, recorded on November, while maximum temperature was 44.8 °C, registered on July (Fig. 1). This figure also shows a strong monthly rainfall variation. After planting, and until January, there was no precipitation, so that two irrigations were carried out to ensure a good germination and seedling emergence. Cumulative rainfall was around 218 mm to which were added the quantities of 20 and 25 mm brought by both irrigations. The rainiest month was March with about 52 mm (Fig. 1). On the other hand, at ATZ, minimum (0.7 °C) and maximum (46.2 °C) temperatures were registered on February and July, respectively (Fig. 2). There was a clear monthly rainfall variation, and cumulative rainfall was around 213.9 mm to which were added the quantity of 30 mm brought by irrigation in late April to have unstressed

Table 2. Analysis of variance (mean_squares) for quantitative traits in M₂ mutant of *Brassica napus* L. generated by EMS, gamma rays and their combination (Treatments) and evaluated in two environments (Locations).

Source of variation	Degree of freedom	Days to flowering	Days to maturity	Plant height (cm)	Number of branches/plant	Number of pods/plant	Number of seeds/pod	Pod length	Pod diameter	1000-seed weight
Treatment	11	500.7***	721.85***	2653.01***	19.02*	909779.77***	122.92***	432.3***	0.62***	2.59***
Location	1	19293.37***	165999.56***	72520.2***	582.95***	8896092.01***	53.11*	27.62	15.45***	11.49***
Treatment *Location	11	87.92**	262.06**	732.68	8.05	718134.03***	22.46*	84.5*	0.23	1.15**

*, **, ***: Significant at 5%, 1% and 1% levels, respectively.

11 degrees of freedom because we have 12 overall treatments (11 mutagenic treatments + one check treatment).

flowering conditions. Like DYT, the rainiest month in ATZ was March, with about 45.5 mm. In both experimental stations, the overall water supply (cumulative rainfall and irrigations) remained much lower than average rainfall, indicating that experiment was conducted under relatively dry conditions.

2.3 Parameters measured

From each treatment, ten mutant plants, for each mutagenic treatment, were taken randomly to study morphological and agronomic parameters in M₂ population. Wild type of the variety “INRA-CZH2” was used as a check. Days to flowering and to maturity of each mutant were calculated as the sum of days from emergence date to date when 50% of plants of this mutant have flowered and matured, respectively. At maturity, plant height (cm), number of branches per plant and number of pods per plant were determined. After harvest, number of seeds per pod was counted in laboratory. Pod length and diameter (mm) were determined using a caliper whilst 1000-seed weight (g) was determined by a precision balance.

2.4 Statistical analysis

Analysis of variance (ANOVA) of gathered data was performed to test significant differences among treatments, environments (sites) and their interaction levels. Duncan’s new multiple range test (DMRT) was applied to compare treatment means. Statistical analysis was conducted with the software package SPSS for Windows (Version 22).

3 Results

According to results of analysis of variance, mutagen treatment (gamma rays, EMS and their combination) significantly affected the variation of all quantitative traits studied in M₂ population (Tab. 2). In addition, there were significant differences between the two INRA Experimental Station of Douyet (DYT) and Sidi Alal Tazi (ATZ) for all parameters with the exception of pod length (Tab. 2). Also, the effect of treatment × environment interaction was significant on these parameters, except plant height, number of branches per plant and pod diameter (Tab. 2). Therefore, the use of mutagen treatment in rapeseed germplasm, regardless of its type, allowed inducing a novel genetic variability through both environments.

Variation in the parameters measured in M₂ progenies, according to the investigated mutagenic treatments is shown in Table 3 for DYT environment, Table 4 for ATZ environment and Table 5 for combined environments. Figure 3 illustrates the novel genetic diversity induced and observed through both experimental environments.

In general, EMS treatments (mainly EMS1-6 and EMS1-7) were found to be more effective for inducing earliness in flowering and maturity, compared to the check and the other mutagenic treatments. In fact, at DYT, mutant lines developed through EMS1-6 and EMS1-7 needed shorter time duration to bloom (99 and 93 days, respectively) and to mature (153 and 171 days, respectively), compared to the check wild variety with 107 and 178 days, respectively (Tab. 3). In ATZ, days to flowering of mutant lines derived from EMS1-6 and EMS1-7 were 76 and 78 days, respectively, and days to maturity were 107 and 106, respectively, whilst the check registered 85 and 111 days, respectively (Tab. 4). Over both locations, it was found that a mutant line derived from EMS1-7 had the lowest average number of days to flowering, 85, and a mutant line coming from EMS1-6 exhibited the lowest average number of days to maturity, 130, compared to 96 and 144 days, respectively, for the check (Tab. 5). On the other hand, seed treatment using 1300 Gy of gamma rays resulted in a slight and non-significant increase in days to flowering and days to maturity over both locations, while combined treatment, through 1300 Gy and 0.8% EMS, induced earliness in flowering and maturity, compared to the check variety (Tab. 5). However, the observed earliness was less pronounced than that induced by EMS1-6 and EMS1-7.

Significant variation was observed on plant height for both locations. At DYT, mutants from 1300 Gy treatment and the check variety had the highest plants, with an average of 156.8 and 155.5 cm, respectively, while mutants developed from combined treatment had the shortest ones, with a mean value of 72.5 cm (Tab. 3). At ATZ, mutant lines coming from 1300 Gy of gamma rays were characterized by the highest plants, with an average of 125 cm, whilst combined treatment and EMS1.4-14 mutants produced the shortest plants, with a mean value of 72.5 and 73.33 cm, respectively (Tab. 4). For combined locations conditions, mutants developed from 1300 Gy exhibited the highest plants (141.74 cm), while there was a decrease trend in average plant height with all other EMS treatments. For EMS1.2-6 and EMS1.2-14, plant height was reduced significantly to 112.55 and 111 cm, respectively, as

Table 3. Effect of different concentrations/doses of chemical mutagen (EMS) and physical mutagen (Gamma rays) and combined mutagen treatment in M₂ generation on quantitative traits in rapeseed evaluated in the Experimental Station of Douyet (DYT).

Treatments	Days to flowering	Days to maturity	Plant height (cm)	Number of branches / plant	Number of pods / plant	Number of seeds / pod	Pod length	Pod diameter	1000-seed weight
Control	107 ^{bcd}	177.6 ^{ab}	155.5 ^a	10.8 ^a	627.9 ^b	28.78 ^a	65.06 ^{abc}	4.28 ^a	2.59 ^{bc}
EMS1-6	98.5 ^{ef}	152.7 ^c	145.4 ^{ab}	9.6 ^a	433.5 ^b	25.93 ^{abc}	63.35 ^{abcd}	3.93 ^{abcd}	2.71 ^{bc}
EMS1-7	92.70 ^f	171.2 ^b	154.8 ^a	12.2 ^a	1504.6 ^a	27.97 ^{ab}	68.8 ^a	4.25 ^{ab}	2.34 ^c
EMS1-14	115 ^a	186 ^{ab}	127.67 ^b	8.17 ^{ab}	496 ^b	19.22 ^d	52.55 ^f	3.62 ^d	1.36 ^d
EMS1.2-6	108.6 ^{abc}	177 ^{ab}	136.5 ^{ab}	8.7 ^{ab}	436.2 ^b	21.77 ^{cd}	54.82 ^{ef}	3.96 ^{abcd}	2.08 ^c
EMS1.2-7	97.7 ^{ef}	177.7 ^{ab}	152 ^a	10.9 ^a	1457.6 ^a	29.19 ^a	67.9 ^{ab}	4.3 ^a	2.26 ^c
EMS1.2-14	112.5 ^{ab}	183.8 ^{ab}	136 ^{ab}	9.8 ^a	457.7 ^b	18.99 ^d	50.88 ^f	3.79 ^{abcd}	2.41 ^c
EMS1.4-6	110.4 ^{ab}	181.6 ^{ab}	150.8 ^a	11 ^a	772.9 ^b	23.89 ^{bc}	60.61 ^{cde}	4.13 ^{abc}	2.39 ^c
EMS1.4-14	102.71 ^{cde}	187 ^a	140.14 ^{ab}	12.14 ^a	505.29 ^b	19.18 ^d	52.41 ^f	3.83 ^{abcd}	3.63 ^a
EMS1.6-6	113.1 ^{ab}	185.2 ^{ab}	154 ^a	10.2 ^a	253.6 ^{bc}	21.83 ^{cd}	56.38 ^{def}	3.95 ^{abcd}	2.65 ^{bc}
1300 Gy	112.2 ^{ab}	184.7 ^{ab}	156.8 ^a	10.6 ^a	459.4 ^b	22.87 ^{cd}	61.19 ^{bcd}	3.91 ^{abcd}	3.24 ^{ab}
CombT	100 ^{de}	177 ^{ab}	72.5 ^c	5.5 ^b	112 ^b	22.65 ^{cd}	56.33 ^{def}	3.96 ^{cd}	2.51 ^c

Concentrations/doses of treatments used are explained and described in [Table 1](#).

Values with different alphabetical superscripts are significantly different ($p \leq 0.05$) according to DMRT.

Table 4. Effect of different concentrations/doses of chemical mutagen (EMS) and physical mutagen (Gamma rays) and combined mutagen treatment in M₂ generation on quantitative traits in rapeseed evaluated in the Experimental Station of Sidi Allal Tazi (ATZ).

Treatments	Days to flowering	Days to maturity	Plant height (cm)	Number of branches / plant	Number of pods / plant	Number of seeds / pod	Pod length (mm)	Pod diameter (mm)	1000-seed weight
Control	84.6 ^{ab}	111 ^{bcd}	111.8 ^{ab}	5.5 ^a	191.5 ^{abc}	24.35 ^{ab}	61.91 ^{abc}	3.78 ^a	1.62 ^{cde}
EMS1-6	75.8 ^d	107.1 ^{cd}	109.7 ^{ab}	6.5 ^a	186.2 ^{abcd}	24.7 ^a	63.64 ^{ab}	3.45 ^{ab}	1.86 ^{bcd}
EMS1-7	77.4 ^{cd}	105.8 ^d	92.9 ^{abc}	7.1 ^a	212.4 ^a	23.31 ^{abc}	60.41 ^{abc}	3.45 ^{ab}	2.45 ^{abc}
EMS1-14	89 ^a	121 ^a	95 ^{abc}	5.33 ^a	102.33 ^{bcd}	22.39 ^{abc}	62.89 ^{ab}	2.74 ^d	1.16 ^e
EMS1.2-6	85.3 ^{ab}	115.6 ^{ab}	88.6 ^{bc}	3.9 ^a	117.5 ^{abcd}	22.77 ^{abc}	58.3 ^{abc}	3.45 ^{ab}	1.62 ^{cde}
EMS1.2-7	82 ^{abcd}	110.2 ^{bcd}	91.5 ^{bc}	7.3 ^a	203.6 ^{ab}	24.92 ^a	64.62 ^a	3.28 ^{bc}	2.28 ^{abcd}
EMS1.2-14	88.4 ^a	120 ^a	86 ^{bc}	5.3 ^a	101.7 ^{bcd}	20.1 ^{cd}	53.47 ^c	3.51 ^{ab}	1.36 ^{de}
EMS1.4-6	84.3 ^{abc}	110.8 ^{bcd}	113 ^{ab}	6.7 ^a	135.1 ^{abcd}	22.34 ^{abc}	59.57 ^{abc}	3.59 ^{ab}	2.15 ^{abcd}
EMS1.4-14	84 ^{abc}	114 ^{abc}	73.33 ^c	5.33 ^a	91 ^{cd}	17.64 ^d	42.7 ^d	2.97 ^{bc}	1.55 ^{cde}
EMS1.6-6	84.7 ^{ab}	115.1 ^{ab}	98.7 ^{abc}	7.6 ^a	151.8 ^{abcd}	20.3 ^{bcd}	57.42 ^{abc}	3.6 ^{ab}	1.95 ^{abcde}
1300 Gy	84 ^{abc}	110 ^{bcd}	125 ^a	7.56 ^a	186.56 ^{abcd}	22.12 ^{abc}	59.98 ^{abc}	3.34 ^{abc}	2.89 ^a
CombT	80.5 ^{bcd}	108 ^{bcd}	72.5 ^c	4.5 ^a	82.5 ^d	23.17 ^{abc}	55.16 ^{bc}	2.9 ^{cd}	2.65 ^{ab}

Concentrations/doses of treatments used are explained and described in [Table 1](#).

Values with different alphabetical superscripts are significantly different ($p \leq 0.05$) according to DMRT.

compared to check (133.65 cm), and for combined treatment, the mutants developed showed the shortest plants, 72.5 cm ([Tab. 5](#)).

At DYT, EMS treatments EMS1-7 and EMS1.4-14 enabled to produce mutants with the most elevated number of branches per plant (12.2 and 12.14, respectively), whereas combined treatment led to mutants with lowest branching (5.5). The check variety had a mean value of 10.8 ([Tab. 3](#)). At ATZ, mutants coming from EMS1.6-6 and 1300 Gy produced the highest number of branches per plant (7.6 and 7.56, respectively), whilst mutants derived from EMS1.2-6 and combined treatment showed the lowest branching (3.9 and 4.5, respectively). The check had an average of 5.5 ([Tab. 4](#)). In

combined environments conditions, the highest average number of branches per plant was 10.10, observed for EMS1.4-14, followed by 9.65 for EMS1-7 and 9.16 for 1300 Gy. The control had a mean value of 8.15. Significant decrease in branching was noticed only for the combined treatment, compared to the check ([Tab. 5](#)).

In each experimental location, number of pods per plant varied significantly. At DYT, mutants derived from EMS1-7 and EMS1.2-7 produced much higher number of pods per plant (1505 and 1458, respectively) than the check, having an average of 628 pods per plant. On the other hand, combined treatment induced a mutant with the lowest number of pods per plant (112) ([Tab. 3](#)). At ATZ, highest number of pods per

Table 5. Effect of different concentrations/doses of chemical mutagen (EMS) and physical mutagen (Gamma rays) and combined mutagen treatment in M₂ generation on quantitative traits in rapeseed evaluated in two different environments (DYT) and (ATZ).

Treatments	Days to flowering	Days to maturity	Plant height (cm)	Number of branches / plant	Number of pods / plant	Number of seeds / pod	Pod length (mm)	Pod diameter (mm)	1000-seed weight
Control	95.80 ^b	144.30 ^{bcd}	133.65 ^{ab}	8.15 ^{abc}	409.7 ^b	26.56 ^a	63.49 ^{ab}	4.03 ^a	2.12 ^{bc}
EMS1-6	87.15 ^{cd}	129.90 ^c	127.55 ^{abc}	8.05 ^{abc}	309.9 ^b	25.31 ^{abc}	63.49 ^{ab}	3.69 ^b	2.28 ^{bc}
EMS1-7	85.00 ^d	138.50 ^d	123.85 ^{abc}	9.65 ^{ab}	858.5 ^a	25.64 ^{ab}	64.61 ^{ab}	3.85 ^{ab}	2.39 ^{bc}
EMS1-14	106.33 ^a	164.33 ^a	116.78 ^{bc}	7.22 ^{bcd}	364.8 ^b	20.28 ^{def}	55.99 ^{cd}	3.33 ^c	1.29 ^d
EMS1.2-6	96.95 ^b	146.30 ^{bcd}	112.55 ^c	6.30 ^{cd}	276.9 ^b	22.27 ^{de}	56.56 ^{cd}	3.71 ^{ab}	1.85 ^c
EMS1.2-7	89.85 ^c	143.95 ^{bcd}	121.75 ^{bc}	8.10 ^{abc}	830.6 ^a	27.05 ^a	66.26 ^a	3.79 ^{ab}	2.27 ^{bc}
EMS1.2-14	100.45 ^b	151.90 ^b	111.00 ^c	8.55 ^{abc}	279.7 ^b	19.55 ^{ef}	52.17 ^{de}	3.65 ^b	1.88 ^c
EMS1.4-6	97.35 ^b	146.20 ^{bcd}	131.90 ^{ab}	8.85 ^{abc}	454.0 ^b	23.11 ^{bcd}	60.09 ^{bc}	3.86 ^{ab}	2.27 ^{bc}
EMS1.4-14	97.10 ^b	165.10 ^a	120.10 ^{bc}	10.10 ^a	381.0 ^b	18.72 ^f	49.49 ^e	3.57 ^{bc}	3.00 ^a
EMS1.6-6	98.90 ^b	150.15 ^{bc}	126.35 ^{abc}	8.90 ^{abc}	202.7 ^b	21.07 ^{def}	56.90 ^{cd}	3.77 ^{ab}	2.29 ^{bc}
1300 Gy	98.84 ^b	149.32 ^{bc}	141.74 ^a	9.16 ^{ab}	330.16 ^b	22.51 ^{cde}	60.62 ^{bc}	3.64 ^b	3.07 ^a
CombT	90.25 ^c	142.5 ^{cd}	72.5 ^d	5 ^d	122.25 ^b	22.91 ^{bcd}	55.74 ^{cd}	3.33 ^c	2.58 ^{ab}

Concentrations/doses of treatments used are explained and described in Table 1.

Values with different alphabetical superscripts are significantly different ($p \leq 0.05$) according to DMRT.



Fig. 3. Novel genetic variability induced by gamma rays and EMS in rapeseed. a: field experiment conducted at Douyet; b: field experiment conducted at Sidi Allal Tazi (2015).

plant (213) was observed in an EMS1-7 mutant, which was significantly higher than that of check (192 pods per plant), whilst the lowest number of pods per plant (83) was found in a mutant developed by combined treatment (Tab. 4). Over both locations, all mutagen treatments affected significantly this trait and, particularly, by applying a treatment of EMS1-7 and EMS1.2-7, a substantial rise in number of pods per plant was observed. In fact, this parameter was 859 and 831 pods in respective mutants, which were more than twice of the check (410 pods). For combined treatment, a mutant with the lowest value (122 pods) was found, when compared to the check and the other mutagenic treatments.

A large variation was noted in number of seeds per pod in both locations. At DYT, the highest mean value was

29.19 seeds/pod, recorded for EMS1.2-7, which remains, however, comparable to the check (28.78 seeds/pod). The lowest mean value was 18.99 seeds/pod, recorded in EMS1.2-14, followed by 19.18 and 19.22 seeds/pod noticed in EMS1.4-14 and EMS1-14, respectively (Tab. 3). At ATZ, EMS1.4-1 induced a mutant having the lowest number of seeds per pod (17.64), compared to the check (24.35 seeds/pod) (Tab. 4). Over both locations, significant decrease in number of seeds per pod was found for gamma rays-300 Gy and combined treatment (22.51 and 22.91 seeds/pod, respectively), when compared to the check (26.6 seeds/pod). Additionally, our data evidenced high levels of EMS treatment affected negatively and drastically this trait. In fact, by using EMS1.2-14 and EMS1.4-14 treatments, number of seeds per plant was reduced to 19.6 and 18.7, respectively (Tab. 5).

Pod length and diameter varied significantly according to mutagenic treatments in all studied environments. At DYT, EMS1-7 enabled obtaining the longest pod (68.8 mm) followed by EMS1.2-7 (67.9 mm), whereas EMS1.2-14 and EMS1.4-14 led to the production of mutants characterized by the shortest pods (50.88 and 52.41 mm, respectively). The check variety had a mean value of 65.06 mm (Tab. 3). At ATZ, the longest pod (64.62 mm) was observed in EMS1.2-7 mutant, whilst the shortest one (42.7 mm) was recorded in EMS1.4-14 mutant. The check had an average of 61.91 mm (Tab. 4). Over both locations, the longest pod, 66.3 mm, was obtained in EMS1.2-7, which was significantly higher than the control average (63.4 mm), whereas the shortest pod, 49.49 mm, was found in EMS1.4-14 (Tab. 5). Regarding pod diameter, it was negatively affected by all mutagenic treatments, and the lowest value ever observed was 3.3 mm, as a result of EMS1-14 and combined treatments, when compared to 4.03 mm recorded in the check (Tab. 5).

Regarding 1000-seed weight, significant variation was observed for all mutagenic treatments and over both locations. At DYT, the highest 1000-seed weight was 3.63 g, recorded in EMS1.4-14 treatment, followed by 3.24 g, registered in gamma

rays-1300 Gy, whilst the lowest one, 1.36 g, was observed for EMS1-14 treatment. The check had a mean value of 2.59 g (Tab. 3). At ATZ, gamma rays-1300 Gy treatment induced the highest 1000-seed weight (2.89 g), while EMS1-14 mutant had the lowest one (1.16 g). The check wild variety had an average of 1.62 g (Tab. 4). In combined location conditions, gamma rays-1300 Gy and EMS1.4-14 treatments enabled to get mutants with highest 1000-seed weight, namely 3.07 and 3 g, respectively, compared to the check (2.12 g) and all other treatments applied in this study (Tab. 5).

4 Discussion

Like as previous findings (Siddiqui *et al.*, 2009; Emrani *et al.*, 2012; More and Malode, 2016), our results confirmed that gamma rays, Ethyl Methane Sulphonate (EMS) and combination of both were efficient mutagenic treatments for increasing genetic variability in rapeseed quantitative traits. It is common approach in various plant breeding programs to use either physical mutagenesis, particularly through gamma radiation, or chemical mutagenesis, mainly by EMS treatment, as a tool to increase and induce novel genetic variability in germplasm. Usefulness of such mutagen treatments is assessed by their mutagenic effectiveness and efficiency (Begum and Dasgupta, 2010). Chemical mutagens are responsible of DNA base substitution by DNA base alkylation (Cooper *et al.*, 2008), whilst ionizing radiation could cause oxidative damage like base modifications and single or double strand breaks (Roldan-Arjona and Ariza, 2009).

In our investigation, mutants derived from low EMS concentrations and short application duration were earlier to flowering and maturity than the original variety. Application of EMS1-6 and EMS1-7 induced earliness in flowering in M₂ mutants, by reducing days to flowering by 9.38% and 11.45%, respectively, compared to wild type. In previous study, Thurling and Depittayan (1992) found one M₃ rapeseed mutant derived from seeds treated with 0.75% EMS during 12 h that flowered 20 days earlier than the parental line. Also, Emrani *et al.* (2012) had found 6 days earliness in flowering time of M₃ lines induced by gamma radiation-1200 Gy treatment. Also, when compared to the wild type, application of EMS1-6 and EMS1-7 induced 14 and 6 days earliness in maturity, respectively, in M₂ mutants. In other words, there is in these mutants a genetic gain in terms of maturity earliness of 9.73 and 4.17%, respectively. In *Brassica juncea*, characterized by much longer crop cycle than *B. napus*, Barve *et al.* (2009) found one mutant derived from seeds treated with 0.02% EMS during 3 h, with 47 days earliness in maturity compared to the check, which indicated a cycle reduction of 33.57%. Early flowering provides sufficient time for seed filling, which could result in better seed yield under short-season and lower rainfall environments. Furthermore, modification of *Brassica* species flowering time is very relevant in agriculture since it may allow extending the geographical range of these crops (Rae *et al.*, 1999). Induction of maturity earliness is one of the traits most frequently altered in mutation breeding programs in *Brassica* crops (Kharkwal *et al.*, 2004), and many early mutants have been developed (Rahman *et al.*, 1992; Das *et al.*, 1999; Barve *et al.*, 2009; Malek and Monshi, 2009). In the present study, compared to wild variety and

mutants coming from the other mutagenic treatments, mutants derived from EMS1-7 were earlier to flowering and maturity, and were also characterized by a higher number of pods per plant through mutation generations and in both environments. According to obtained data over both experimental locations and to the plant architecture of EMS1-7, we would expect more stability and adaptability of this mutant over various and contrasted environments. In addition, this mutant was generally more adapted than check and other mutants to stressful environments associated with low rainfall, high temperature and contrasted sowing dates. In fact, experimental conditions in both environments were characterized by high maximum average temperatures during March-April period coinciding with flowering and early seed filling (Figs. 1 and 2). Therefore, one could expect that early flowering mutant would be more performant in terms of seed yield and seed oil content under such conditions.

As a result of using high concentration of EMS during long time and combination of physical and chemical mutagens, a significant decrease in plant height and stature was noticed, as compared to control. In a previous study, Kumar and Yadav (2010) had also reported that plant height was found to be significantly reduced by high doses of mutagenic treatment, and in some cases, plants responded positively to lower mutagen doses and recorded a slight increase in their height. Dwarfism in mutant plants can be explained by decline of mitotic activity of meristematic tissues and reduction in moisture content in seeds (Khalil *et al.*, 1986). Improvement in seed yield may be associated with reduction in plant height (More and Malode, 2016), and one could remember that use of dwarfing genes was a key factor in the success of green revolution (Khush, 2001). Dwarfing genes may improve seed yield through reduced lodging and increasing harvest index. Many mutation breeders did isolate dwarf mutants in rapeseed and mustard using physical and chemical mutagenesis (Chauhan and Kumar, 1986; Rai and Singh, 1993; Shah *et al.*, 1999; Javed *et al.*, 2003; Zeng *et al.*, 2011; Wei *et al.*, 2018). On the other hand, an increase in plant height was observed as a result of seed irradiation with gamma rays-1300 Gy. This is in perfect agreement with findings of Emrani *et al.* (2012) who reported the increase in this trait in M₂ and M₃ generation mutants induced by gamma rays ionizing.

In the same way, an increase of 1000-weight was observed in mutant plants derived from seeds treated by gamma rays-1300 Gy. Similar results were reported by Siddiqui *et al.* (2009) having used 1000 Gy of gamma rays to ionize their rapeseed seeds. Also, mutants in oilseed *Brassica* with improved seed size were obtained using gamma rays in other studies (Chauhan and Kumar 1986; Shah *et al.*, 1990). In our work, one could observe that overall 1000-seed weight, regardless of the mutagenic treatments, was lower than standard and common value which is about 3.50–4.00 g (Nabloussi, 2015). This was likely due to unfavorable environmental conditions under which the experiment was conducted, particularly characterized by drought and heat stress in both experimental environments. In fact, it is well known that seed weight is an important seed yield component that is strongly affected by environmental conditions (Diepenbrock, 2000).

Following the use of gamma rays and EMS mutagens, a novel genetic variability was induced and promising mutants

were obtained. However, EMS was more interesting as it generated the most performant mutants in terms of flowering earliness and seed yield, mainly when it was applied in low dose and during moderate time. Previous reports had also shown that EMS effectiveness was higher than gamma rays (Thakur and Sethi, 1995; Kharkwal, 1998; Solanki, 2005; Wani, 2009; Begum and Dasgupta, 2010).

However, acquisition of novel desirable mutations has been, recently, facilitated by the introduction of targeted genome editing (GE) technologies, especially clustered regularly interspaced short palindromic repeats (CRISPR)/ (CRISPR)-associated 9 (Cas9) (Braatz *et al.*, 2017). The application of CRISPR-Cas9-targeted mutagenesis in plants has been reported not only in *Arabidopsis* (Fauser *et al.*, 2014; Feng *et al.*, 2014) but also in crops like wheat (Wang *et al.*, 2014), tomato (Brooks *et al.*, 2014), and rice (Li *et al.*, 2016). In rapeseed, some few studies have already presented targeted genome editing mediated by the CRISPR/Cas9 system (Braatz *et al.*, 2017; Okuzaki *et al.*, 2018; Sun *et al.*, 2018). Therefore, as more and more genes are identified for their function, mutagenesis in rapeseed plant breeding could be managed by more efficient ways than chemical/physical and random ways.

5 Conclusion

In conclusion, we confirmed that gamma rays, Ethyl Methane Sulphonate (EMS) and combination of both were potent and highly effective for inducing novel variability in some important agronomic traits in rapeseed. Different mutagenic treatments enabled to develop various mutants with modified and desirable characteristics. Interestingly, modifications and particular characteristics of these mutants were recorded at the level of M_1 and M_2 plants, and were maintained over both environments of this study. This suggested that these mutants were stable and thus, they could be used and exploited efficiently in rapeseed breeding program. Among the various mutagenic treatments used in present investigation, EMS1-7 was found to be more effective in obtaining earliness in flowering and maturity and to increase number of pods per plant, which is the most important component of seed yield. On the other hand, mutants derived from high dose of EMS during long time and from combination treatment of physical and chemical mutagens decreased significantly plant height and, as a result, short statured plants were obtained. Furthermore, plants derived from seeds irradiated by gamma rays-1300 Gy exhibited the highest 1000-seed weight. Overall, the mutants developed in this study will increase the available genetic variability in Moroccan rapeseed germplasm. Many of these mutants might be stable and thus may enable to develop efficiently and quickly new rapeseed cultivars with different desirable traits.

References

Ali HMA, Shah SA. 2013. Evaluation and selection of rapeseed (*Brassica napus* L.) mutant lines for yield performance using augmented design. *J Anim Plant Sci* 23: 1125–1130.

Barve YY, Gupta RK, Bhadauria SS, Thakre RP, Pawar SE. 2009. Induced mutations for development of *B. juncea* canola quality varieties suitable for Indian agro-climatic conditions. In: Induced

plant mutations in genomics era. Food and Agriculture Organization of the United States, pp. 373–375.

Begum T, Dasgupta T. 2010. A comparison of the effects of physical and chemical mutagens in sesame (*Sesamum indicum* L.). *Genet Mol Biol* 33: 761–766.

Braatz J, Harloff HJ, Mascher M, Stein N, Himmelbach A, Jung C. 2017. CRISPR-Cas9 targeted mutagenesis leads to simultaneous modification of different homoeologous gene copies in polyploid oilseed rape (*Brassica napus*). *Plant Physiol* 174(2): 935–942.

Brooks C, Nekrasov V, Lippman ZB, Van Eck J. 2014. Efficient gene editing in tomato in the first generation using the clustered regularly interspaced short palindromic repeats/CRISPR-associated9 system. *Plant Physiol* 166: 1292–1297.

Bus A, Körber N, Snowdon RJ, Stich B. 2011. Patterns of molecular variation in a species-wide germplasm set of *Brassica napus*. *Theor Appl Genet* 123(8): 1413–1423.

Chauhan YS, Kumar K. 1986. Gamma ray induced chocolate seeded mutant in *Brassica campestris* var. Yellow Sarson. *Curr Sci* 55: 410.

Cooper JL, Till BJ, Laport RG, *et al.* 2008. TILLING to detect induced mutations in soybean. *BMC Plant Biol* 8: 9.

Das ML, Rahman A, Malek MA. 1999. Two early maturing and high yielding varieties of rapeseed developed through induced mutation technique. *Bang J Bot* 28: 27–33.

Diepenbrock W. 2000. Yield analysis of winter oilseed rape (*Brassica napus* L.): A review 2000. *Field Crops Res* 67: 35–47.

Emrani SN, Arzani A, Saeidi G, *et al.* 2012. Evaluation of induced genetic variability in agronomic traits by gamma irradiation in canola (*Brassica napus* L.). *Pak J Bot* 44: 1281–1288.

Emrani SN, Harloff H, Gudi O, Kopisch Obuch F, Jung C. 2015. Reduction in sinapine content in rapeseed (*Brassica napus* L.) by induced mutations in sinapine biosynthesis genes. *Mol Breed* 35 (1): 37.

FAOSTAT. 2018. Available from <http://www.fao.org/faostat/en/#data/> (last consult: 2018/12/04).

Fauser F, Schiml S, Puchta H. 2014. Both CRISPR/Cas-based nucleases and nickases can be used efficiently for genome engineering in *Arabidopsis thaliana*. *Plant J* 79: 348–359.

Feng Z, Mao Y, Xu N, *et al.* 2014. Multigeneration analysis reveals the inheritance, specificity, and patterns of CRISPR/Cas-induced gene modifications in *Arabidopsis*. *Proc Natl Acad Sci USA* 111: 4632–4637.

Ferrie AMR, Taylor DC, MacKenzie SL, Rakow G, Raney JP, Keller WA. 2008. Microspore mutagenesis of *Brassica* species for fatty acid modifications: a preliminary evaluation. *Plant Breed* 127: 501–506.

Hasan M, Seyis F, Badani AG, *et al.* 2006. Analysis of genetic diversity in the *Brassica napus* L. gene pool using SSR markers. *Genet Resour Crop Evolut* 53(4): 793–802.

Hussain S, Khan WM, Khan MS, *et al.* 2017. Mutagenic effect of sodium azide (NaN_3) on M_2 generation of *Brassica napus* L. (variety Dunkled). *Pure Appl Biol* 6: 226–236.

Javed MA, Siddiqui MA, Khan MKR, *et al.* 2003. Development of high yielding mutants of *Brassica campestris* L. cv. Toria selection through gamma rays irradiation. *Asian J Plant Sci* 2: 192–195.

Khalil SJ, Rehman S, Afridi K, Jan MT. 1986. Damage induced by gamma irradiation in morphological and chemical characteristics of barley. *Sarhad J Agric* 2: 45–54.

Kharkwal MC. 1998. Induced mutations in chickpea (*Cicer arietinum* L.). I. Comparative mutagenic effectiveness and efficiency of physical and chemical mutagens. *Indian J Genet* 58: 159–167.

Kharkwal MC, Pandey RN, Pawar SE. 2004. Mutation breeding for crop improvement. In: Jain HK, Kharkwal MC, eds. Mendelian to

- molecular approaches. New Delhi, India: Narosa Publishing House, pp: 601–645.
- Khush GS. 2001. Green revolution: The way forward. *Nat Rev Genet* 2: 815–822.
- Kumar G, Yadav RS. 2010. EMS induced genetic disorders in sesame (*Sesamum indurum* L.). *Rom J Biol Plant Biol* 55: 97–104.
- Lee YH, Park W, Kim KS, *et al.* 2018. EMS-induced mutation of an endoplasmic reticulum oleate desaturase gene (FAD2-2) results in elevated oleic acid content in rapeseed (*Brassica napus* L.). *Euphytica* 214: 28.
- Li T, Yaokui L, Dan Z, *et al.* 2016. Characteristic and inheritance analysis of targeted mutagenesis mediated by genome editing in rice. *Hereditas (Beijing)* 38(8): 746–755.
- Malek MA, Monshi FI. 2009. Performance evaluation of rapeseed mutants. *Bang J Agric Res* 36: 81–84.
- Meinke DW, Cherry JM, Dean C, Rounsley SD, Koornneef M. 1998. *Arabidopsis thaliana*: A model plant for genome analysis. *Science* 282: 679–682.
- More UA, Malode SN. 2016. Mutagenic effect of EMS on quantitative characters of *Brassica napus* L. Cv. Excel in M₁ generation. *J Global Biosci* 5: 4018–4025.
- Nabloussi A. 2015. Amélioration génétique du colza: enjeux et réalisations pour un développement durable de la filière. Rabat, Maroc: INRA-DIC Edition. ISBN: 9789954-593-27-1.
- Okuzaki A, Ogawa T, Koizuka C, *et al.* 2018. CRISPR/Cas9-mediated genome editing of the fatty acid desaturase 2 gene in *Brassica napus*. *Plant Physiol Biochem* 131: 63–69.
- Parry MA, Madgwick PJ, Bayon C, *et al.* 2009. Mutation discovery for crop improvement. *J Exp Bot* 60: 2817–2825.
- Rae AM, Howell EC, Kearsey MJ. 1999. More QTL for flowering time revealed by substitution lines in *Brassica oleracea*. *Heredity* 83: 586–596.
- Rahman A. 1990. Evolution of improved varieties of rapeseed mustard and sesame through induced mutations. In: *Proceedings on "Mutations breeding of oilseed crops" FAO/IAEA, Vienna, Austria*, pp. 57–67.
- Rahman A, Das ML, Pathan AJ. 1992. New high yielding mutant varieties of mustard (*Brassica campestris* L., var. Yellow Sarson). *J Nucl Agric Biol* 21: 281–285.
- Rai B, Singh D. 1993. A note on potential sources of dwarfing genes in Indian rapeseed (*Brassica campestris* L. Prain). *Ind J Genet* 53: 153–156.
- Roldan-Arjona T, Ariza RR. 2009. Repair and tolerance of oxidative DNA damage in plants. *Mutat Res* 681: 169–179.
- Schnurbush T, Mollers C, Becker HC. 2000. A mutant of *B. napus* with increased palmitic acid content. *Plant Breed* 119: 141–144.
- Shah SA, Ali I, Rahman K. 1990. Induction and selection of superior genetic variables of oilseed rape (*Brassica napus* L.). *Nucl* 27: 37–40.
- Shah SA, Ali I, Iqbal MM, Khattak SU, Rahman K. 1999. Evolution of high yielding and early flowering variety of rapeseed (*Brassica napus* L.) through in-vivo mutagenesis. In: *Proceedings of 3rd International Symposium New Genetical Approaches to Crop Improvement-III Nuclear Institute of Agriculture, Tandojam, Pakistan*, pp. 47–53.
- Sharafi Y, Majidi MM, Goli SH, Rashidi F. 2015. Oil content and fatty acids composition in *Brassica* species. *Int J Food Prop* 18: 2145–2154.
- Siddiqui MA, Khan IA, Khatri A. 2009. Induced quantitative variability by gamma rays and ethylmethane sulphonate alone and in combination in rapeseed (*Brassica napus* L.). *Pak J Bot* 41: 1189–1195.
- Singh BD. 2000. Mutations in crop improvement. In: *Plant breed. Principles and methods*. New Delhi, Kalyani Publishers, pp. 598–631.
- Solanki IS. 2005. Isolation of macromutations and mutagenic effectiveness and efficiency in lentil (*Lens culinaris* Medik). *Indian J Genet* 65: 264–268.
- Spasibonek S. 2006. New mutants of winter rapeseed (*B. napus* L.) with changed fatty acid composition. *Plant Breed* 125: 259–267.
- Sun Q, Lin L, Liu D, *et al.* 2018. CRISPR/Cas9-mediated multiplex genome editing of the BnWRKY11 and BnWRKY70 genes in *Brassica napus* L. *Int J Mol Sci* 19(9): 2716.
- Thakur JR, Sethi GS. 1995. Comparative mutagenicity of gamma rays, ethyl methane sulphonate and sodium azide in barley (*Hordeum vulgare* L.). *Crop Res* 9: 350–357.
- Thurling N, Depittayanan V. 1992. EMS induction of early flowering mutants in spring rape (*Brassica napus*). *Plant Breed* 108: 177–184.
- Tshilenge-Lukanda L, Kalonji-Mbuyi A, Nkongolo KK, Kizungu RV. 2013. Effect of gamma irradiation on morpho-agronomic characteristics of groundnut (*Arachis hypogaea* L.). *Am J Plant Sci* 04: 2186–2192.
- Van Harten AM. 1998. Mutation breeding, theory and practical applications. Cambridge, United Kingdom: Cambridge University Press, pp. 127–140.
- Velasco L, Fernandez-martinez JM, De Haro A. 2008. Inheritance of reduced linolenic acid content in the Ethiopian mustard mutant N2-4961. *Plant Breed* 127: 263–265.
- Wanasundara JPD, McIntosh TC, Perera SP, Withana-Gamage TS, Mitra P. 2016. Canola/rapeseed protein-functionality and nutrition. *OCL* 23(4): D407.
- Wang Y, Cheng X, Shan Q, *et al.* 2014. Simultaneous editing of three homoeoalleles in hexaploid bread wheat confers heritable resistance to powdery mildew. *Nat Biotechnol* 32: 947–951.
- Wani AA. 2009. Mutagenic effectiveness and efficiency of gamma rays, ethyl methane sulphonate and their combination treatments in chickpea (*Cicer arietinum* L.). *Asian J Plant Sci* 8: 318–321.
- Wei C, Zhu L, Wen J, *et al.* 2018. Morphological, transcriptomics and biochemical characterization of new dwarf mutant of *Brassica napus*. *Plant Sci* 270: 97–113.
- Zeng X, Zhu L, Chen Y, *et al.* 2011. Identification, fine mapping and characterization of a dwarf mutant (bnaC. dwarf) in *Brassica napus*. *Theor Appl Genet* 122(2): 421–428.

Cite this article as: Channaoui S, Labhilili M, Mouhib M, Mazouz H, El Fechtali M, Nabloussi A. 2019. Development and evaluation of diverse promising rapeseed (*Brassica napus* L.) mutants using physical and chemical mutagens. *OCL* 26: 35.