

*LIPIDS AND BRAIN*  
*LIPIDES ET CERVEAU*

## Phospholipid, arachidonate and eicosanoid signaling in schizophrenia

Erik Messamore<sup>1</sup> and Jeffrey K. Yao<sup>2,3,4,\*</sup>

<sup>1</sup> University of Cincinnati and Lindner Center for Hope, Cincinnati, OH, USA

<sup>2</sup> VA Pittsburgh Healthcare System, Pittsburgh, PA 15240, USA

<sup>3</sup> Department of Psychiatry, Western Psychiatric Institute Clinic, University of Pittsburgh Medical Center, Pittsburgh, PA 15213, USA

<sup>4</sup> Department of Pharmaceutical Sciences, University of Pittsburgh School of Pharmacy, Pittsburgh, PA 15213, USA

Received 16 September 2015 – Accepted 21 September 2015

**Abstract** – This paper reviews the potential role of arachidonic acid in the pathophysiology of schizophrenia. We discuss how abnormal levels of arachidonic acid may arise, and how dysregulation of signaling molecules derived from it have the potential to disrupt not only dopamine signaling, but numerous other physiological processes associated with the illness. Pharmacological doses of niacin stimulate the release of arachidonic acid; and arachidonic acid-derived molecules in turn dilate blood vessels in the skin. A blunted skin flush response to niacin is reliably observed among patients with schizophrenia. The niacin response abnormality may thus serve as a biomarker to identify a physiological subtype of schizophrenia associated with defective arachidonic acid-derived signaling.

**Keywords:** Phospholipids / arachidonic acid / eicosanoids / niacin-induced flushing, endophenotype marker / schizophrenia

**Résumé** – **Signalisation des phospholipides, de l'arachidonate et de l'éicosanoïde dans la schizophrénie.** Cet article examine le rôle potentiel de l'acide arachidonique dans la physiopathologie de la schizophrénie. Il est discuté comment des niveaux anormaux d'acide arachidonique peuvent survenir, et comment la dérégulation des molécules de signalisation qui en découle est capable de perturber non seulement la signalisation de la dopamine, mais aussi de nombreux autres processus physiologiques associés avec cette maladie. Des doses pharmacologiques de niacine stimulent la libération d'acide arachidonique; des molécules dérivées de l'acide arachidonique dilatent à leur tour les vaisseaux sanguins dans la peau. Une brusque rougeur de la peau en réponse à la niacine est observée de manière constante parmi les patients atteints de schizophrénie. La réponse anormale à la niacine peut donc servir de biomarqueur afin d'identifier un sous-type physiologique de la schizophrénie, associé à un système défectueux de signalisation des dérivés de l'acide arachidonique.

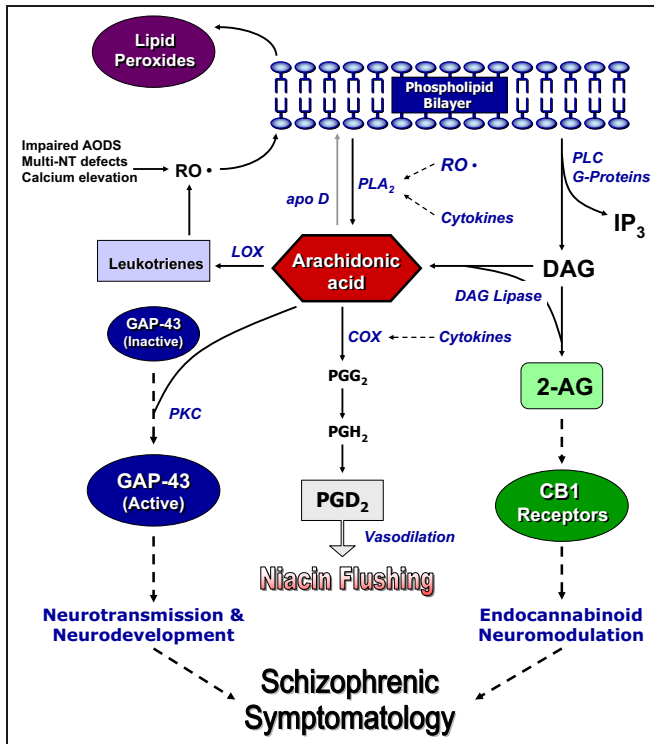
### 1 Introduction

Schizophrenia (SZ) is a complex behavioral syndrome associated with diverse biochemical and physiological abnormalities. This paper will describe how arachidonic acid (AA) and abnormalities related to its metabolism may to some extent unify some seemingly unrelated biochemical findings in SZ (Horobin, 1998; Skosnik and Yao, 2003). We will also describe how a blunted skin flush response to niacin may serve as a biomarker for AA-related signaling defects, and suggest that AA-related abnormalities may represent a distinct physiological subtype within the SZ syndrome (Messamore, 2003; Messamore *et al.*, 2010; Yao *et al.*, 2015).

\* Correspondence: [jkyao@pitt.edu](mailto:jkyao@pitt.edu)

### 2 Heterogeneity of the Schizophrenias

Despite early acknowledgement of the likelihood that SZ is not a unitary illness (Bleuler, 1920), for most of the 20th century it was usually studied as if it were a single disease entity. The dominant view was that of a unitary illness with differing modes of expression. This view of SZ, however, is no longer tenable. The variable therapeutic response to antipsychotic medication provides strong evidence against a unitary cause of SZ (Garver *et al.*, 2000). In fact, the preponderance of evidence is consistent with SZ having numerous, etiologically distinct causes (Jablensky, 2006). Efforts toward understanding its various causes, and toward developing improved, etiologically-focused treatments, may be more fruitful if the schizophrenias were deconstructed into physiologically-based



**Fig. 1.** An overview of phospholipids turnover, arachidonic acid, and eicosanoid signaling in schizophrenia symptomatology (adapted from Skosnik and Yao, 2003). Abbreviations: AODS, antioxidant defense system; NT, neurotransmitters; RO, reactive oxygen species; apoD, apolipoprotein D; PLA<sub>2</sub>, phospholipase A<sub>2</sub>; PLC, phospholipase C; LOX, lipoxygenase; AA, arachidonic acid; DAG, diacylglycerol; COX, cyclooxygenase; 2-AG, 2-arachidonoyl glycerol; GAP, growth-associated protein; PKC, phosphokinase C; PGG<sub>2</sub>, prostaglandin G<sub>2</sub>; PGH<sub>2</sub>, prostaglandin H<sub>2</sub>; PGD<sub>2</sub>, prostaglandin D<sub>2</sub>; CB, cannabinoid.

intermediate phenotypes (Braff *et al.*, 2007; Keshavan *et al.*, 2008). We and others have observed evidence suggesting that one of these intermediate phenotypes may involve phospholipid signaling abnormalities that prominently involve dysregulation of AA levels or the actions of its physiologically active metabolites.

### 3 Phospholipid, arachidonate and eicosanoid (PAE) signaling in SZ

A phospholipid turnover abnormality is present in many cases of SZ. Disordered phospholipid turnover with resulting changes in the levels and metabolic destinations of AA can unite seemingly unrelated neurochemical and clinical observations in SZ.

A model relating phospholipid signaling to SZ-relevant neurochemical abnormalities can be found in Figure 1. As shown, several factors – including oxidative stress and cytokine release – have the potential to lyse membrane phospholipids, resulting in the release of AA and, over time, reduce its cell membrane levels. Overactivity of phospholipase A<sub>2</sub> (PLA<sub>2</sub>) or phospholipase C (PLC) enzymes would deplete the membrane-lysable pool of AA. In turn, changes in

the availability of AA and AA-derived signaling molecules affect the release or circulating levels of several neuroactive molecules, including: GAP-43; the neurotransmitters dopamine and glutamate, and the endocannabinoids anandamide and 2-arachidonoylglycerol (2-AG). Alterations in AA may also affect the inflammatory response, which can then affect PLA<sub>2</sub> release *via* cytokines, further exacerbating phospholipid turnover and AA release. These diverse disruptions have the potential to impact many neuronal signaling pathways relevant to psychosis (Horrobin, 1998; Peet *et al.*, 1994; Skosnik & Yao, 2003).

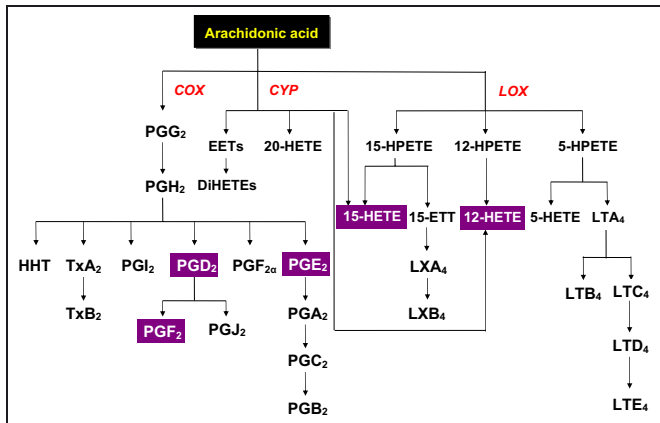
Low levels of AA have been observed postmortem in the cerebral cortex of SZ patients (Horrobin *et al.*, 1991; Yao *et al.*, 2000). Low AA levels have been identified in red blood cell (RBC) membranes from patients with chronic SZ (Glen *et al.*, 1994; Peet *et al.*, 1994; Vaddadi *et al.*, 1986; Yao *et al.*, 1994a), as well as first-episode neuroleptic-naïve SZ (Avrindakshan *et al.*, 2003; Reddy *et al.*, 2004). Thus, abnormally low AA levels have been observed in brain as well as RBC membrane phospholipids from patients with SZ (further reviewed by Conklin *et al.*, 2007; Peet, 2007; Skosnik and Yao, 2003; Yao, 2003). These changes could conceivably lead to decreased synthesis of eicosanoids. Collectively, these changes can account for numerous physiological and clinical observations in SZ.

Utilizing varying types of samples (*e.g.* plasma, RBC, platelets, postmortem brain, etc.) and methodologies (<sup>31</sup>P Magnetic resonance spectroscopy, platelet function, niacin-induced flushing, etc.), somewhat consistent patterns of decreased polyunsaturated fatty acids (PUFAs) and increased phospholipid turnover are apparent (Bentsen *et al.*, 2011; Horrobin, 1998; Mahadik and Yao, 2006; Peet, 2007; Pettegrew *et al.*, 1991; Skosnik and Yao, 2003; Yao *et al.*, 1994a), particularly in relation to AA.

In addition to the formation of second messengers, AA released from membrane phospholipids can be converted to a variety of biologically active metabolites, collectively termed eicosanoids, through the concerted reactions of cyclooxygenase (COX), lipoxygenase (LOX) and cytochrome P-450 (CYP). Eicosanoids can modulate neural cell function, and can mediate several pathophysiological processes (Bazan, 2006). Since AA is the major C20 polyunsaturated fatty acid (PUFA) in mammalian tissues, the prostaglandin-2 (PG2) and thromboxanes-2 (TX2) series are the predominant classes of eicosanoids (Fig. 2). Inhibition of COX by nonsteroidal anti-inflammatory drugs has revealed the significance of PG2 in the regulation of nerve conduction, neurotransmitter release, inflammation, pain, fever, immune responses, apoptosis and psychosis (Akhondzadeh *et al.*, 2007).

### 4 Clinical and physiological effects of AA and eicosanoids

AA deficiency, or abnormal regulation of AA-derived signaling could explain several clinical and physiological findings in SZ. Association between low RBC membrane AA levels and the expression of the negative symptom syndrome (apathy, social withdrawal, affective flattening, etc.) has been reported (Glen *et al.*, 1994). In other studies, low RBC



**Fig. 2.** Arachidonic acid cascade (adapted from Condray and Yao, 2011). EET, PGI<sub>2</sub>, PGD<sub>2</sub>, PGE<sub>2</sub>, PGF<sub>2α</sub>, 12-HETE and 15-HETE are considered as potential vasodilators.

membrane levels of dihomo- $\gamma$ -linolenic acid, the immediate precursor of AA, were associated with increased psychosis ratings (Yao *et al.*, 1994b) and disorganized symptoms (Doris *et al.*, 1998). Among SZ patients receiving fatty acid supplementation therapy, increased RBC AA contents were associated with a reduction in the severity of symptoms (Peet *et al.*, 2002).

Aspirin inhibits vascular dilation during electrical stimulation of cerebral cortex slices *in vitro* (Zonta *et al.*, 2003); and pharmacological inhibition of COX-2, or COX-2 gene knock-out, reduces by 40 to 50% the cerebral blood flow response of the sensory cortex to whisker stimulation in mice (Niwa *et al.*, 2000). Thus, an eicosanoid signaling abnormality could potentially contribute to the cerebral neurovascular coupling defects that have been repeatedly observed in SZ (Berman *et al.*, 1992; Liddle *et al.*, 1992; Weinberger *et al.*, 1992).

Many signs and symptoms of SZ are suggestive of excess dopamine activity (Snyder, 1981). Prostaglandin D<sub>2</sub> (PGD<sub>2</sub>) and other PGs stimulate the production of cAMP and thereby exert functional antagonism of dopamine-D<sub>2</sub> receptors, which modulate inhibition of cAMP synthesis (Ono *et al.*, 1992; Wanibuchi and Usuda, 1990). PGD<sub>2</sub> therefore counteracts the biochemical and behavioral effects of dopamine *in vitro* and *in vivo*. PGD<sub>2</sub> administration blocks the behavioral effects of dopaminergic agents such as apomorphine, L-DOPA, and amphetamine; and induces catalepsy in a manner identical to dopamine-D<sub>2</sub> receptor antagonists such as haloperidol (Ono *et al.*, 1992; Wanibuchi and Usuda, 1990). Deficient PGD<sub>2</sub> signaling would thus be expected to produce the signs of excessive dopamine activity that are seen in SZ. A role for PGs as endogenous anti-dopaminergic agents is supported by the findings that inhibitors of PG synthesis can cause psychotic symptoms in some people and can worsen psychotic symptoms in SZ patients (Hoppman *et al.*, 1991; Sussman and Magid, 2000). The relationship between SZ and COX products is complex, however, as there are also reports suggesting that COX inhibitors may augment the efficacy of antipsychotic medications (Akhondzadeh *et al.*, 2007; Laan *et al.*, 2010; Müller *et al.*, 2002). These seemingly contradictory findings nonetheless speak to the relevance of prostaglandins to the pathophysiology of the schizophrenias while also pointing toward their etiologic diversity.

biology of the schizophrenias while also pointing toward their etiologic diversity.

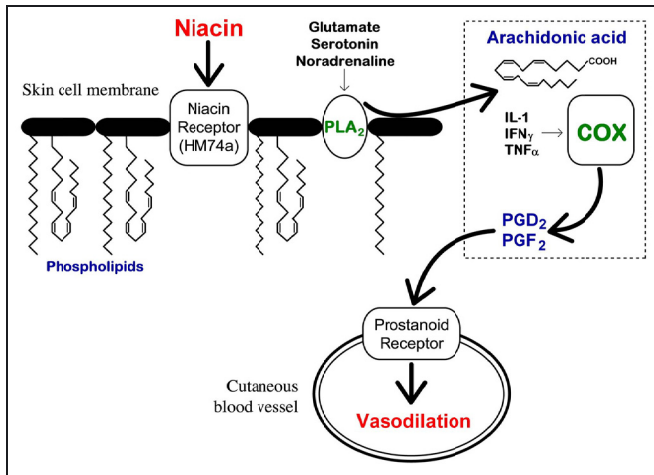
A deficiency of prostaglandins has previously been related to SZ (Horrobin, 1977). It is likely that reduced AA availability may in part explain a variety of clinical observations that are usually ignored by the receptor-based etiological hypotheses (Horrobin, 1998). For example, in SZ, there appears to be a lower risk of arthritis and other inflammatory diseases (Eaton *et al.*, 1992), greater resistance to pain (Davis *et al.*, 1979, Horrobin, 1977), and remission of psychosis during fever has been observed (Horrobin, 1977). These effects could be secondary to reduced eicosanoid signaling.

## 5 Niacin Flush response as a test for eicosanoid abnormalities

Niacin (nicotinic acid), at sufficient doses, is well-known to dilate blood vessels in the skin resulting in a visible skin flush. The biochemical mechanism (Fig. 3) of this response has been studied over the course of several decades so many of the key steps have been well described. Niacin-induced skin flushing is prominently mediated by metabolites of AA. Thus, it may serve as a physiological marker of the integrity of phospholipid-arachidonate-eicosanoid (PAE) signaling. We will now describe how the niacin response abnormality might define a physiologically distinct subtype of SZ.

## 6 Mediation of niacin flush by AA metabolites

The levels of PGD<sub>2</sub> and 9 $\alpha$ ,11 $\beta$ -PGF<sub>2</sub> (a stable metabolite of PGD<sub>2</sub>) rise markedly in skin exudate or venous blood following oral or topical administration of niacin in humans (Eklund *et al.*, 1979; Kobza Black *et al.*, 1982; Morrow *et al.*, 1992). Evidence favors PGD<sub>2</sub> as the predominant mediator of the flush response (Morrow *et al.*, 1989). However PGE<sub>2</sub> levels also increase in response to niacin (Eklund *et al.*, 1979; Kobza Black *et al.*, 1982). In man, the flush response to niacin can be completely abolished by COX inhibitors such as aspirin or indomethacin (Svedmyr *et al.*, 1977; Wilkin *et al.*, 1982). In mice, the flush response to niacin can be eliminated by knocking out genes for either the niacin receptor or COX-1, consistent with an exclusive and obligatory role for this pathway in niacin-evoked skin flushing (Benyó *et al.* 2005). Classical vasodilatory mediators, such as histamine, bradykinin, serotonin, and acetylcholine do undergo changes in their skin transudate levels after niacin challenge (Morrow *et al.*, 1992; Plummer *et al.*, 1977). However, Papaliadis *et al.* (2008) showed that niacin induced the release of serotonin from human platelets *in vitro* and elevated blood levels of serotonin after intraperitoneal injection of niacin in rats. Although further study will be needed to clarify the possible role of serotonin, the vast majority of evidence points to a strongly predominant role of AA metabolites as primary mediators of the flush response to niacin. Other eicosanoid vasodilators, however, have not been well studied in SZ.



**Fig. 3.** The mechanism of niacin-induced skin flushing (adapted from Messamore *et al.*, 2010). Abbreviations: COX, cyclooxygenase; PLA<sub>2</sub>, phospholipase A<sub>2</sub>; IL, interleukin; INF, interferon; TNF, tumor necrosis factor; PGD<sub>2</sub>, prostaglandin D<sub>2</sub>; PGF<sub>2</sub>, prostaglandin F<sub>2</sub>.

## 7 Mechanism of the niacin flush response

A model of our current understanding of the mechanism of niacin-induced skin flushing is illustrated in Figure 3. Niacin interacts with a specific G-protein coupled receptor, HM74a, located on dermal macrophages and adipocytes (Benyó *et al.*, 2006; Urade *et al.*, 1989); its activation by niacin stimulates PLA<sub>2</sub> mediated formation of AA (Tang *et al.*, 2006). Formation of AA is the rate-limiting step in the biosynthesis of the vasodilatory PGD<sub>2</sub> and E<sub>2</sub> (PGE<sub>2</sub>) (Murakami and Kudo 2004). These prostaglandins bind to specific prostanoid receptors on vascular smooth muscle within the skin. Activation of prostanoid receptors dilates cutaneous blood vessels (Lai *et al.*, 2007), and a visible skin flush arises from the ensuing increased blood flow (Benyó *et al.*, 2005; Maciejewski-Lenoir *et al.*, 2006; Morrow *et al.*, 1989, 1992).

## 8 The abnormal niacin response in SZ

Abrams Hoffer first reported observations that a substantial portion of patients with SZ are unusually resistant to the skin flush effect of pharmacologically-dosed niacin (Hoffer, 1962). Citing this work, and supporting these observations with clinical evidence of prostaglandin defects in the illness, Horrobin (1980a, 1980b) proposed that the niacin response abnormality could be used to demonstrate a prostaglandin signaling abnormality in SZ. Abnormally attenuated skin flushing in response to niacin has been very widely replicated in samples of patients with SZ (Messamore, 2003; Smesny *et al.*, 2003; Yao *et al.*, 2015). The niacin response abnormality is consistently over-represented in SZ compared to both healthy control groups (Messamore, 2003), or psychiatrically ill comparison groups with major depression (Bosveld-van Haandel *et al.*, 2006) or bipolar disorder (Liu *et al.*, 2007; Ross *et al.*, 2004; Yao *et al.*, 2015).

Consistent with the view that SZ is an etiologically heterogeneous disorder, the niacin response abnormality is not

present in all patients with SZ. The prevalence of the niacin abnormality was pegged at 80 to 90% in some studies (Puri *et al.*, 2001; Ward *et al.*, 1998). However, we have observed a prevalence of about 30% in SZ patients in two separate experiments, and 20% in first-degree relatives of patients with SZ (unpublished data). There has been very little consistency across reports with respect to the methods to stimulate or measure blood flow. This methodological variability accounts for the disparity of estimates of the prevalence of the niacin response abnormality in SZ. If, however, the same methods are used, there is remarkable consistency in the ability to find higher rates of abnormality among SZ patients *versus* other clinical or healthy comparison groups.

## 9 Biochemical correlates of abnormal niacin response

Lower RBC membrane AA levels are associated with decreased niacin sensitivity, but this expected correlation has only been described in a healthy control group (Messamore *et al.*, 2010). Although lower AA levels were found in patients who did not flush in response to oral niacin (Glen *et al.*, 1996), no correlation between AA levels and niacin response was observed in two different groups of patients with SZ (Maclean *et al.*, 2003; Messamore *et al.*, 2010). On the other hand, an unexpected correlation was observed between maximal niacin-induced blood flow and red blood cell membrane levels of adrenic acid (22:4 n-6), the elongation product of AA. Adrenic acid has vasodilatory actions in bovine coronary arteries and in arteries of the cortical layer of the adrenal gland (Kop *et al.*, 2010). Lack of correlation between AA levels and niacin response may result from a homeostatic imbalance within the n-6 PUFAs pathway in SZ. The significance of adrenic acid levels as they relate to niacin-induced blood flow response deserves further attention.

RBC membrane levels AA are normally tightly correlated with its immediate elongation product adrenic acid (22:4 n-6). However, this expected correlation is abolished in SZ (Messamore *et al.*, 2010). Similar precursor-product dysregulations have also shown in tryptophan (Yao *et al.*, 2010b) and purine (Yao *et al.*, 2010a) pathways. Although many of these correlated relationships persist across disease or medication status, others are lost among patients with SZ.

## 10 Clinical correlates of abnormal niacin response

The extent to which abnormal niacin-induced skin flushing associates with clinical manifestations of SZ has recently been reviewed (Messamore, 2012). Impaired niacin-induced flushing has been associated with greater severity of both positive and negative symptoms of SZ (Berger *et al.*, 2002; Glen *et al.*, 1996; Hudson *et al.*, 1997; Smesny *et al.*, 2003). Niacin response impairment was significantly linked to inorganic phosphate levels revealed by <sup>31</sup>P magnetic resonance spectroscopy (Puri *et al.*, 2007). This suggests that niacin-abnormal SZ patients may have higher levels of cerebral energy metabolism. Niacin response, as reflected in the 'volumetric niacin response

score' appears much more impaired among SZ patients with a history of violence (Puri *et al.*, 2007) compared to ordinary SZ patients (Puri *et al.*, 2002). Niacin response impairment is associated with worsened global functioning among patients with SZ (Messamore, 2012). Additionally, the niacin response abnormality is associated with significant cognitive impairment in SZ (Nilsson *et al.*, 2015).

## 11 The niacin response abnormality is not an artifact of medication or smoking status

Evidence suggests that neither antipsychotic drugs nor smoking significantly affect the niacin skin flush response (Mills *et al.* 1997; Shah *et al.* 2000; Smesny *et al.* 2001; Turenne *et al.* 2001). The niacin skin flush is not affected by local anesthetics, suggesting that local neurotransmitter release is not involved in niacin-induced vasodilatation (Winkleman *et al.*, 1965). There is no correlation between antipsychotic drug dose and niacin sensitivity in patients with SZ (Hudson *et al.*, 1997; Messamore, 2003). Neither has a significant difference in niacin sensitivity been found between medicated versus unmedicated patients (Shah *et al.*, 2000). In contrast, bipolar disorder patients who take antipsychotic medications have a normal or even enhanced flush response (Hudson *et al.*, 1997). Moreover, niacin subsensitivity occurs in first-degree relatives of schizophrenics, which suggests that this is a heritable trait, independent of medication status (Shah *et al.*, 1999; Waldo, 1999). On the other hand, Tavares *et al.* (2003) reported that 4 out of 13 SZ patients with niacin subsensitivity became sensitive to niacin after 8 weeks of atypical antipsychotic drug treatment. If such drugs do tend to normalize niacin sensitivity, then the potential bias in patient sample would be to underestimate the prevalence or magnitude of the niacin response abnormality in SZ. Taken together, comparing the niacin-induced flushing between medicated and unmedicated SZ patients, or unmedicated, non-psychotic relatives, suggests that the niacin response abnormality in SZ is not an artifact of antipsychotic medications (Chang *et al.*, 2009; Lin *et al.*, 2007; Maclean *et al.*, 2003; Shah *et al.*, 2000).

Similarly, it appears that nicotine use has no effect on the niacin-induced flushing response (Chang *et al.*, 2009; Liu *et al.*, 2007; Messamore, 2003, 2010; Ross *et al.*, 2004; Shah *et al.*, 2000; Smesny *et al.*, 2003). Our recent findings (Yao *et al.*, 2015) also support the notion that niacin sensitivity is not significantly affected by smoking.

## 12 Possible mechanisms for the niacin response abnormality in SZ

As depicted in Figure 3, there are several possible explanations for the finding of niacin subsensitivity in SZ. These include: abnormal signaling at the niacin receptor; abnormal presentation of free AA to COX; abnormal COX activity; abnormal conversion of initial COX products to vasodilatory end products; or abnormal signaling at prostanoid receptors in vascular smooth muscle. Support for abnormal niacin receptor

signaling is provided by a report of decreased niacin receptor expression in postmortem brain from SZ patients (Miller and Dulay, 2008).

Presentation of free AA to COX is accomplished by the action of PLA<sub>2</sub>, which may be abnormally active in niacin-subsensitive SZ (Hudson *et al.*, 1996; Tavares *et al.*, 2003). Any process leading to diminished output of prostaglandins could potentially account for niacin subsensitivity. Abnormal COX activity in SZ was indirectly demonstrated by Das and Khan (1998).

Cyclooxygenase action on AA initially produces prostaglandin H<sub>2</sub>, which is subsequently converted by tissue-specific isomerases to vasodilatory end-products. Isomerase abnormalities have been detected in discrete brain regions from patients with a variety of mental illnesses (Maida *et al.*, 2006).

It is intriguing to note that none of these pathways is specifically targeted by existing antipsychotic medications. Discovering the mechanisms responsible for the niacin response abnormality may inform the development of novel treatment strategies. Such treatments would likely involve targets outside the traditional monoamine receptors and thus would be expected to augment the efficacy of current medications. Elucidating the mechanism of the niacin response abnormality may also lead to physiologically-informed categorization of mental illness.

## 13 Conclusions

There is a compelling need to categorize the schizophrenias according to physiological criteria. Ideally such physiologically-defined categories would lead more easily than the older, traditional schema to insights about biochemical or genetic causes – and would suggest more tailored and effective treatments. Although this goal is desirable, there have been relatively few leads as to which physiological abnormalities may be more promising to follow.

We suggest that the niacin skin flush response abnormality is a viable candidate for the physiological subtyping of SZ. In contrast to the incompletely-understood etiology of SZ, the mechanism of niacin-induced skin flushing in man is relatively well-characterized. This detailed knowledge, coupled with the accessibility of skin for scientific study, presents a technically feasible, straightforward, and economically attractive opportunity to broaden our understanding of the biochemical changes that accompany SZ.

It is intriguing to note that none of the biochemical pathways mediating the niacin response is specifically targeted by existing antipsychotic medications. Discovering the mechanisms responsible for the niacin response abnormality may inform the development of novel treatment strategies. Such treatments would likely involve targets outside the traditional monoamine receptors and thus would be expected to augment the efficacy of current medications. As the metabolic and genetic underpinnings of this biomarker are elucidated, it should yield valuable insights into the complex pathophysiology of schizophrenic illness.

## Disclosure

The authors declare no conflict of interest

**Acknowledgements.** Supported in part by Department of Veterans Affairs [Merit Reviews 1I01CX000110 (JKY) and Senior Research Career Scientist Award (JKY)], and the VA Pittsburgh Healthcare System. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. The contents of this article do not represent the views of the Department of Veterans Affairs or the United States Government.

## References

- Akhondzadeh S, Tabatabaee M, Amini H, *et al.* 2007. Celecoxib as adjunctive therapy in schizophrenia: a double-blind, randomized and placebo-controlled trial. *Schizophr. Res.* 90: 179–185.
- Arvindakshan M, Sitasawad S, Debsikdar V, *et al.* 2003. Essential polyunsaturated fatty acid and lipid peroxide levels in never-medicated and medicated schizophrenia patients. *Biol. Psychiatry* 53: 56–64.
- Bazan NG. 2006. Eicosanoids, docosanoids, platelet-activating factor and inflammation. In: Siegel GJ, Albers WR, Brady ST, Price DL, eds., *Basic neurochemistry: Molecular, cellular, and medical aspects*. 7<sup>th</sup> ed., Burlington, MA: Academic Press, pp. 575–591.
- Bentsen H, Solberg DK, Refsum H, *et al.* 2011. Bimodal distribution of polyunsaturated fatty acids in schizophrenia suggests two endophenotypes of the disorder. *Biol. Psychiatry* 70: 97–105
- Benyó Z, Gille Z, Kero J, Csiky M, *et al.* 2005. GPR109A (PUMA-G/HM74A) mediates nicotinic acid – induced flushing. *J. Clin. Invest.* 115: 3634–3640.
- Benyó Z, Gille Z, Bennett CL, Clausen BE, Offermanns S. 2006. Nicotinic acid-induced flushing is mediated by activation of epidermal langerhans cells. *Mol. Pharmacol.* 70: 1844–1849.
- Berger G, Yuen H, McGorry P. 2002. The topical niacin flush test in early psychosis (Abstract). *Schizophr. Res.* 53: S38.
- Berman KF, Torrey EF, Daniel DG, Weinberger DR. 1992. Regional cerebral blood flow in monozygotic twins discordant and concordant for schizophrenia. *Arch. Gen. Psychiatry* 49: 927–934.
- Bleuler E. 1920. *Lehrbuch der Psychiatrie*, Berlin: Springer reprinted English translation (1976) Textbook of psychiatry. New York: Arno Press.
- Bosveld-van Haandel L, Knegtering R, Kluiters H, van den Bosch RJ 2006. Niacin skin flushing in schizophrenic and depressed patients and healthy controls. *Psychiatry Res.* 143: 303–306.
- Braff DL, Freedman R, Schork NJ, Gottesman II. 2007. Deconstructing schizophrenia: an overview of the use of endophenotypes in order to understand a complex disorder. *Schizophr. Bull.* 33: 21–32.
- Chang, S.S., Liu, C.M., Lin, S.H., *et al.* 2009. Impaired flush response to niacin skin patch among schizophrenia patients and their nonpsychotic relatives: the effect of genetic loading. *Schizophr Bull* 35: 213–221.
- Condray R, Yao JK. 2011 Cognition, dopamine and bioactive lipids in schizophrenia. *Front. Biosci.* S3: 298–330.
- Conklin SM, Muldoon MF, Reddy RD, Yao JK. 2007. Fatty acids and psychiatric disorders. In: Chow C.K. ed., *Fatty acids in foods and their health implications*, 3rd ed. New York: Marcel Dekker, Inc., pp. 1229–1256.
- Das I, Khan NS. 1998. Increased arachidonic acid induced platelet chemiluminescence indicates cyclooxygenase overactivity in schizophrenic subjects. *Prostagland. Leukot. Essent. Fatty Acids* 58: 165–168.
- Davis GC, Buchsbaum MS, van Kammen DP, Bunney WE Jr. 1979. Analgesia to pain stimuli in schizophrenics and its reversal by naltrexone. *Psychiatry Res.* 1: 61–69.
- Doris AB, Wahle K, MacDonald A, *et al.* 1998. Red cell membrane fatty acids, cytosolic phospholipase A<sub>2</sub> and schizophrenia. *Schizophr. Res.* 31: 185–196.
- Eaton WW, Hayward C, Ram R. 1992. Schizophrenia and rheumatoid arthritis: a review. *Schizophr. Res.* 6: 181–192.
- Eklund B, Kaijser L, Nowak J, Wennmalm A. 1979. Prostaglandins contribute to the vasodilation induced by nicotinic acid. *Prostaglandins* 17: 821–830.
- Garver DL, Holcomb JA, Christensen JD. 2000. Heterogeneity of response to antipsychotics from multiple disorders in the schizophrenia spectrum. *J. Clin. Psychiatry* 61: 964–972.
- Glen AI, Glen EM, Horrobin DF, *et al.* 1994. A red cell membrane abnormality in a subgroup of schizophrenic patients: evidence for two diseases. *Schizophr. Res.* 12: 53–61.
- Glen AI, Cooper SJ, Rybakowski J, Vaddadi K, Brayshaw N, Horrobin DF. 1996. Membrane fatty acids, niacin flushing and clinical parameters. *Prostagland. Leukotr. Essent. Fatty Acids* 55: 9–15.
- Hoffer A. 1962. *Niacin therapy in psychiatry*. Springfield, IL: Charles Thomas.
- Hoppman RA, Peden JG, Over SK. 1991. Central nervous system side effects of nonsteroidal anti-inflammatory drugs. *Arch. Intern. Med.* 151: 1309–1313.
- Horrobin DF. 1977. Schizophrenia as a prostaglandin deficiency disease. *Lancet* 1: 936–937.
- Horrobin DF. 1980a. Niacin flushing, prostaglandin E and evening primrose oil: a possible objective test for monitoring therapy in schizophrenia. *Orthomol. Psychiatry* 91: 33–34
- Horrobin DF. 1980b. Schizophrenia: a biochemical disorder? *Biomedicine* 32: 54–55.
- Horrobin DF. 1998. The membrane phospholipid hypothesis as a biochemical basis for the neuro-developmental concept of schizophrenia. *Schizophr. Res.* 30: 193–208.
- Horrobin DF, Manku MS, Hillman H, Iain A, Glen M. 1991. Fatty acid levels in the brains of schizophrenics and normal controls. *Biol. Psychiatry* 30: 795–805.
- Hudson CJ, Kennedy JL, Gotowiec A, *et al.* 1996. Genetic variant near cytosolic phospholipase A<sub>2</sub> associated with schizophrenia. *Schizophr. Res.* 21: 111–116.
- Hudson CJ, Lin A, Cogan S, Cashman F, Warsh JJ. 1997. The niacin challenge test: clinical manifestation of altered transmembrane signal transduction in schizophrenia. *Biol. Psychiatry* 41: 507–513.
- Jablensky A. 2006 Subtyping schizophrenia: implications for genetic research. *Mol. Psychiatry* 11: 815–836.
- Keshavan MS, Tandon R, Boutros NN, Nasrallah HA. 2008. Schizophrenia, just the facts: what we know in 2008 Part 3: neurobiology. *Schizophr. Res.* 106: 89–107.
- Khanna N, Altmeyer W Zhuo J, Steven A. 2015 Functional neuroimaging: Fundamental principles and clinical applications. *Neuroradiol. J.* 28: 87–96.
- Kobza Black A, Greaves MW, Hensby CN. 1982. The effect of systemic prednisolone on arachidonic acid, and prostaglandin E<sub>2</sub> and F<sub>2</sub> alpha levels in human cutaneous inflammation. *Br. J. Clin. Pharmacol.* 14: 391–394.
- Kop PG, Zhang D., Gauthier KM, *et al.* 2010. Adrenic acid metabolites as endogenous endothelium-derived and zona glomerulosa-derived hyperpolarizing factors. *Hypertension* 55: 547–554.
- Laan W, Grobbee, D, Selten JP, Heijnen CJ, Kahn RS, Burger H. 2010. Adjuvant aspirin therapy reduces symptoms of schizophrenia spectrum disorders: results from a randomized, double-blind, placebo controlled trial. *J. Clin. Psychiatry* 71, 520–527.

- Lai, E, De Lepeleire, I, Crumley, TM, *et al.* 2007. Suppression of niacin-induced vasodilation with an antagonist to prostaglandin D<sub>2</sub> receptor subtype 1. *Clin. Pharmacol. Ther.* 81: 849–857.
- Liddle PF, Friston KJ, Frith CD, Hirsch SR, Jones T, Frackowiak RS. 1992. Patterns of cerebral blood flow in schizophrenia. *Brit. J. Psychiatry* 160: 179–186.
- Lin SH, Liu CM, Chang SS, *et al.* 2007. Familial aggregation in skin flush response to niacin patch among schizophrenic patients and their nonpsychotic relatives. *Schizophr. Bull.* 33: 174–182.
- Liu CM, Chang SS, Liao SC, *et al.* 2007. Absent response to niacin skin patch is specific to schizophrenia and independent of smoking. *Psychiatr. Res.* 152: 181–187.
- Maciejewski-Lenoir D., Richman JG, Hakak Y, Gaidarov I, Behan DP, Connolly DT. 2006. Langerhans cells release prostaglandin D<sub>2</sub> in response to nicotinic acid. *J. Invest. Dermatol.* 126: 2637–2646.
- Macleay R, Ward PE, Glen I, Roberts SJ, Ross BM, 2003. On the relationship between methylnicotinate-induced skin flush and fatty acids levels in acute psychosis. *Prog. Neuropsychopharmacol. Biol. Psychiatry.* 27: 927–933.
- Mahadik SP, Yao JK. 2006. Phospholipids in schizophrenia. In: Lieberman, J.A., Stroup, T.S., and Perkins, D.O. eds. Textbook of schizophrenia. Washington, DC: The American Psychiatric Publishing, Inc., pp. 117–135.
- Maida ME, Hurley SD, Daeschner JA, Moore AH, O'Banion MK. 2006. Cytosolic prostaglandin E<sub>2</sub> synthase (cPGES) expression is decreased in discrete cortical regions in psychiatric disease. *Brain Res.* 1103: 164–172.
- Messamore E. 2003. Relationship between the niacin skin flush response and essential fatty acids in schizophrenia. *Prostagland. Leukot. Essent. Fatty Acids* 69: 413–419.
- Messamore E. 2012. Niacin subsensitivity is associated with functional impairment in schizophrenia. *Schizophr. Res.* 137: 180–184
- Messamore E, Hoffman WF, Yao JK. 2010. Niacin sensitivity and the arachidonic acid pathway in schizophrenia. *Schizophr. Res.* 122: 248–256.
- Miller, C.L., Dulay, J.R. 2008. The high-affinity niacin receptor HM74A is decreased in the anterior cingulate cortex of individuals with schizophrenia. *Brain Res. Bull.* 77: 33–41.
- Mills CM, Hill SA, Marks R. 1997. Transdermal nicotine suppresses cutaneous inflammation. *Arch. Dermatol.* 133: 823–902
- Morrow JD, Parsons WG, Roberts LJ. 1989. Release of markedly increased quantities of prostaglandin D<sub>2</sub> *in vivo* in humans following the administration of nicotinic acid. *Prostaglandins* 38: 263–274.
- Morrow JD, Awad JA, Oates JA, Roberts LJ. 1992. Identification of skin as a major site of prostaglandin D<sub>2</sub> release following oral administration of niacin in humans. *J. Invest. Dermatol.* 98: 812–815.
- Müller N, Riedel M, Scheppach C, *et al.* 2002. Beneficial antipsychotic effects of celecoxib add-on therapy compared to risperidone alone in schizophrenia. *Am. J. Psychiatry* 159: 1029–1034.
- Murakami M, Kudo I. 2004. Recent advances in molecular biology and physiology of the prostaglandin E<sub>2</sub>-biosynthetic pathway. *Prog. Lipid Res.* 43: 3–35.
- Nilsson BM, Holm G, Hultman CM, Ekselius L. 2015 Cognition and autonomic function in schizophrenia: inferior cognitive test performance in electrodermal and niacin skin flush non-responders. *Eur. Psychiatry* 30: 8–13.
- Niwa K, Araki E, Morham SG, Ross ME, Iadecola C. 2000. Cyclooxygenase-2 contributes to functional hyperemia in whisker-barrel cortex. *J. Neurosci.* 20: 763–770.
- Ono N, Abiru T, Kamiya H. 1992. Influences of cyclooxygenase inhibitors on the cataleptic behavior induced by haloperidol in mice. *Prostagland. Leukotr. Essent. Fatty Acids* 46: 59–63.
- Papaliadis D, Boucher W, Kempuraj D, *et al.* 2008. Niacin-induced “flush” involves release of prostaglandin D<sub>2</sub> from mast cells and serotonin from platelets: Evidence from human cells *in vitro* and an animal model. *J. Pharmacol. Exp. Ther.* 327: 665–672.
- Peet M. 2007. Membrane fatty acid deficit in schizophrenia and mood disorders. In: Reddy R and Yao JK eds. Fatty acids and oxidative stress in neuropsychiatric disorders. New York: Nova Science Publishers, Inc., pp. 101–114.
- Peet M, Laugharne JDE, Horrobin DF, Reynolds GP. 1994. Arachidonic acid: A common link in the biology of schizophrenia? *Arch. Gen. Psychiatry* 51: 665–666.
- Peet M, Horrobin DF, Group EEMS. 2002. A dose ranging exploratory study of the effects of ethyleicosapentaenoate in patients with persistent schizophrenic symptoms. *J. Psychiatr. Res.* 36: 7–18.
- Pettegrew JW, Keshavan MS, Panchalingam K, *et al.* 1991. Alterations in brain high-energy phosphate and membrane phospholipid metabolism in first-episode, drug-naïve schizophrenics. A pilot study of the dorsal prefrontal cortex by *in vivo* phosphorus 31 nuclear magnetic resonance spectroscopy. *Arch. Gen. Psychiatry.* 48: 563–568.
- Plummer NA, Hensby CN, Black AK, Greaves MW. 1977. Prostaglandin activity in sustained inflammation of human skin before and after aspirin. *Clin. Sci. Mol. Med.* 52: 615–620.
- Puri BK, Easton T, Das I, Kidane L, Richardson AJ. 2001. The niacin skin flush test in schizophrenia: a replication study. *Int. J. Clin. Pract.* 55: 368–370.
- Puri BK, Hirsch SR, Easton T, Richardson AJ. 2002. A volumetric biochemical niacin flush based index that noninvasively detects fatty acid deficiency in schizophrenia. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 26: 49–52.
- Puri BK, Richardson AJ, Counsell SJ, *et al.* 2007. Negative correlation between cerebral inorganic phosphate and the volumetric niacin response in male patients with schizophrenia who have seriously and dangerously violently offended: a (31)P magnetic resonance spectroscopy study. *Prostagland. Leukot. Essent. Fatty Acids* 77: 97–99.
- Reddy RD, Keshavan MS, Yao JK. 2004. Reduced red blood cell membrane polyunsaturated fatty acids in first-episode schizophrenia at neuroleptic-naïve baseline. *Schizophr. Bull.* 30: 901–911.
- Ross BM, Hughes B, Turenne S, Seeman M, Warsh JJ. 2004. Reduced vasodilatory response to methyl-nicotinate in schizophrenia as assessed by laser doppler flowmetry. *Eur. Neuropsychopharma* 14: 191–197.
- Shah SH, Ramchand CN, Peet M. 1999. The niacin skin flush test: first-degree relatives show responses intermediate between patients and controls. *Schizophr. Res.* 36: 314.
- Shah SH, Vankar GK, Peet M, Ramchand CN. 2000. Unmedicated schizophrenic patients have a reduced skin flush in response to topical niacin. *Schizophr. Res.* 43: 163–164.
- Skosnik PD, Yao JK. 2003. From phospholipid and fatty acid defects to altered neurotransmission: Is arachidonic acid a nexus in the pathophysiology of schizophrenia? *Prostagland. Leukot. Essent. Fatty Acids* 69: 367–384
- Smesny S, Riemann S, Riehemann S, Bellemann ME, Sauer H. 2001. Quantitative measurement of induced skin reddening using optical reflection spectroscopy—methodology and clinical application. *Biomed. Tech. (Berl).* 46: 280–286.

- Smesny S, Berger G, Rosburg T, *et al.* 2003. Potential use of the topical niacin skin test in early psychosis a combined approach using optical reflection spectroscopy and a descriptive rating scale. *J. Psychiatr. Res.* 37: 237–247.
- Snyder SH. 1981. Dopamine receptors, neuroleptics, and schizophrenia. *Am. J. Psychiatry.* 138: 460–464.
- Sussman N, Magid S. 2000. Psychiatric manifestations of nonsteroidal antiinflammatory drugs. *Primary Psychiatry* 7: 26–30.
- Svedmyr N, Heggelund A, Aberg G, 1977. Influence of indomethacin on flush induced by nicotinic acid in man. *Acta. Pharmacol. Toxicol.* 41: 397–400.
- Tang Y, Zhou L, Gunnet JW, Wines PG, Cryan EV, Demarest KT. 2006. Enhancement of arachidonic acid signaling pathway by nicotinic acid receptor HM74A. *Biochem. Biophys. Res. Commun.* 345: 29–37.
- Tavares H, Yacubian J, Talib LL, Barbosa NR, Gattaz WF. 2003. Increased phospholipase A2 activity in schizophrenia with absent response to niacin. *Schizophr. Res.* 61: 1–6.
- Turenne SD, Seeman M, Ross BM. 2001. An animal model of nicotinic-acid-induced vasodilation: effect of haloperidol, caffeine, and nicotine upon nicotinic acid response *Schizophr. Res.* 50: 191–197
- Urade Y, Ujihara M, Horiguchi Y, Ikai K, Hayaishi O. 1989. The major source of endogenous prostaglandin D<sub>2</sub> production is likely antigen-presenting cells. Localization of glutathione-requiring prostaglandin D synthetase in histiocytes, dendritic, and Kupffer cells in various rat tissues. *J. Immunol.* 143: 2982–2989.
- Vaddadi KS, Gilleard CJ, Mindham RH Butler R. 1986. A controlled trial of prostaglandin E1 precursor in chronic neuroleptic resistant schizophrenic patients. *Psychopharmacol.* 88: 362–367.
- Waldo MC. 1999. Co-distribution of sensory gating and impaired niacin flush response in the parents of schizophrenics. *Schizophr. Res.* 40: 49–53.
- Wanibuchi F, Usuda S. 1990. Synergistic effects between D-1 and D-2 dopamine antagonists on catalepsy in rats. *Psychopharmacology* 102: 339–342.
- Ward PE, Sutherland J, Glen EM, Glen AI. 1998. Niacin skin flush in schizophrenia: a preliminary report. *Schizophr. Res.* 29: 269–274.
- Weinberger DR, Berman KF, Suddath R, Torrey EF. 1992. Evidence of dysfunction of a prefrontal-limbic network in schizophrenia: a magnetic resonance imaging and regional cerebral blood flow study of discordant monozygotic twins. *Am J. Psychiatry.* 149: 890–897.
- Wilkin JK, Wilkin O, Kapp R, Donachie R, Chernosky ME, Buckner J. 1982. Aspirin blocks nicotinic acid-induced flushing. *Clin. Pharmacol. Ther.* 31: 478–482.
- Winklemann RK, Wilhelmj CM, Horner FA. 1965. Experimental studies on dermatographism. *Arch. Dermatol.* 92: 436–442.
- Yao JK. 2003. Abnormalities of fatty acid metabolism in red cells, platelets and brain in schizophrenia. In: Peet M, Glen I, Horrobin DF, eds., *Phospholipid spectrum disorders in psychiatry and neurology*, 2nd ed. Lancashire, UK: Marius Press, pp. 193–212.
- Yao JK, van Kammen DP, Welker JA. 1994a. Red blood cell membrane dynamics in schizophrenia. II. Fatty acid composition. *Schizophr. Res.* 13: 217–226.
- Yao JK, van Kammen DP, Gurklis J. 1994b. Red blood cell membrane dynamics in schizophrenia. III. Correlation of fatty acid abnormalities with clinical measures. *Schizophr. Res.* 13: 227–232.
- Yao JK, Leonard S, Reddy R. 2000. Membrane phospholipid abnormalities in postmortem brains from schizophrenic patients. *Schizophr. Res.* 42: 7–17.
- Yao JK, Dougherty GG, Reddy RD, *et al.* 2010a. Homeostatic imbalance of purine catabolism in first-episode neuroleptic-naïve patients with schizophrenia. *PLoS One* 5: e9508.
- Yao JK, Dougherty GG, Reddy RD, *et al.* 2010b. Altered interactions of tryptophan metabolites in first-episode neuroleptic-naïve patients with schizophrenia. *Mol. Psychiatry.* 15: 938–953.
- Yao JK, Dougherty Jr GG, Gautier CH, *et al.* 2015. Prevalence and specificity of the abnormal niacin response: A potential endophenotype marker in schizophrenia. *Schizophr. Bull.* DOI: [10.1093/schbul/sbv130](https://doi.org/10.1093/schbul/sbv130).
- Zonta M, Angulo MC, Gobbo S, Rosengarten B, Hossmann KA, Pozzan T, Carmignot G. 2003. Neuron-to-astrocyte signaling is central to the dynamic control of brain microcirculation. *Nat. Neurosci.* 6: 43–50.

**Cite this article as:** Erik Messamore, Jeffrey K. Yao. Phospholipid, arachidonate and eicosanoid signaling in schizophrenia. OCL 2016, 23(1) D112.